

SYNTHESIS, CHARACTERIZATION AND LIPOPHILICITY STUDIES OF BENZOTHAIAZOLE DERIVATIVES

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ABSTRACT

Benzothiazole is a heterocyclic chemical compound. The aim is to synthesize title compounds by facile synthesis procedure starting from p- substituted aniline & potassium thiocyanate to give 6-substituted 2-amino benzothiazole. Subsequently added substituted benzaldehyde, in chlorine, O-NO₂, M-NO₂, P-NO₂, and OCH₃ to give these compounds' lipophilicity is determined. Our experiment is to determine lipophilicity as it has become a critical parameter in the pharmaceutical industry. This indicates the relationship of a drug with its biological pharmacokinetic and metabolic properties lipophilicity can be measured by the distribution of a drug between the organic phase which is generally n-octanol pre-saturated with water and the aqueous phase as the water it can be experimentally determined by using various methods like shake flask method, potentiometric, and chromatographic methods. The structure of the synthesized compound was confirmed on the basis of spectral studies (U.V, I.R, H.NMR). The lipophilicity parameter of (4-substituted benzylidene -6- substituted benzo[d] thiazol - 2 amine) was determined by using solvents such as n-octanol and benzene, following the shake flask method and titration method. Which was found that the product is distributed greater in the organic phase than in the aqueous phase, which is lipophilic in nature.

KEYWORDS: Lipophilicity, Synthesis of benzothiazol derivatives, Characterization.

INTRODUCTION

The objective isto improve the knowledge base required for the synthesis, isolation, purification, and characterization of various pharmaceuticals. Heterocyclic chemistry plays a very important role in medicinal chemistry as well as inorganic chemistry. Most of the drug molecules formed and possess therapeutic activity due to the heterocyclic scaffold.^[6-12] A slight change in heterocyclic moiety leads to a major therapeutic change in the drug molecule. Benzothiazole can serve as a unique and versatile moiety for experimental drug design. Benzothiazole and its derivatives are essentially chemical compounds with tremendous application in the research area as well as in pharmaceutical chemistry because of its potent and significant pharmacological activities. Heterocyclic analogs and their derivatives have attracted strong interest in medicinal chemistry due to their biological and pharmacological properties.^[12-15]

Lipophilicity

Lipophilicity is defined as the affinity of a molecule or moiety to a lipophilic environment. It is an important parameter described by the partition coefficient (*P*). It describes the partition of the compound between aquatic and nonpolar solvents. It is commonly assessed by the distribution in a liquid-liquid system (with the shake-flask method) or liquid-solid (using chromatographic techniques). Lipophilicity is a complex effect of molecular interactions both between solute and solvent as well as interactions between solvent molecules in each phase. The substance, when dissolved in the solvent, disrupts its structure, which causes the breakdown of intermolecular bonds. The substance must create a free space in a given volume of solvent.^[15-23] It requires a certain amount of energy and results in a clear correlation of the lipophilicity with the volume or surface of the dissolved molecule.

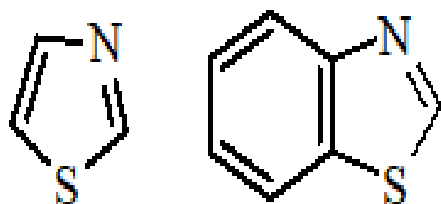


Fig. 1: Structure of benzthiazole derivatives.^[29]

Bioavailability and Lipophilicity

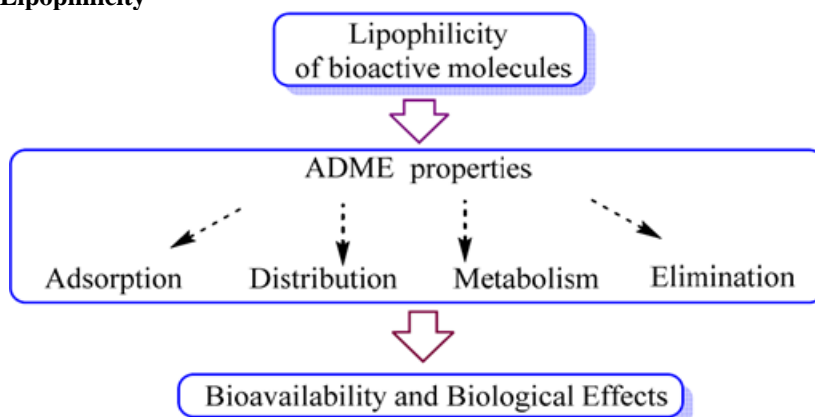


Fig. 2: ADME effects on lipophilicity.^[26]

Bioavailability is a parameter that is highly dependent on solubility, permeability, and clearance; moreover, all three parameters, in turn, depend on lipophilicity. Studies report that the optimum range of lipophilicity to achieve good availability is log P (logarithm of the partition coefficient) between zero and three. Also, parameters, such as rotatable bonds and ionization state are also good predictors of bioavailability as these also have an effect on parameters, including hydrogen bonding, lipophilicity, molecular volume, and ionizability. Thus, there has been an increase in the drive to control properties, such as solubility and lipophilicity, in order to improve the quality and likelihood of therapeutic success of drug compounds compounds.^[30-35]

FORMULA

$$\text{Partition coefficient}(p) = \frac{\text{conc. of Drug in org. phase.}}{\text{conc. of Drug in aq. Phase.}}$$

MATERIALS AND METHODS

Precoated silica gel G plates were used to monitor the progress of reaction as well as to check the purity of the compounds: n-octanol:benzene(1:2), IR spectrometer (model Shimadzu) in the range of 666-4000 cm^{-1} and spectra were interpreted. FTIR spectrometer(model 8400 S) showing different vibration levels by using KBR pellet in the region 3433 cm^{-1} , NMR spectra the samples are analysed on advance 300 MH_2 and spectra were interpreted. Melting point of the organic compounds were determined by capillary tube method.

Experimental Section

First step: General synthesis of 2-amino-6-fluoro-7-chloro-benzothiazole

To glacial acetic acid (20ml) cooled below room temperature were added 8gm (0.08mol) of potassium thiocyanate and 1.45g (0.01 mol) of fluoro chloro aniline. The mixture was cooled in a water bath and mechanically stirred while 1.6ml of bromine in 6ml of glacial acetic acid was added, from a dropping funnel at such a rate that the temperature never rise beyond room temperature. After all the bromine was added (105min),

the solution was stirred for 2 hours below room temperature and at room temperature for 10 hours, it was then allowed to stand overnight, during which period an orange precipitate settle at the bottom, water (6ml) was added quickly and the slurry was heated at 85 $^{\circ}\text{C}$ on a steam bath and filtered hot. The orange residue was placed in a reaction flask and treated with 10ml of glacial acetic acid heated again to 85 $^{\circ}\text{C}$ and filtered hot. The combined filtrate was cooled and neutralized with concentrated ammonia solution to p^{H} 6 A dark yellow precipitate was collected. Recrystallized from benzene, ethanol of (1:1) after treatment with animal charcoal gave yellow plates of 2-amino-6-fluoro-7-chloro-(1,3)-benzothiazole. After drying in an oven at 80 $^{\circ}\text{C}$, the dry material (1gm51.02%) melted at 210-212 $^{\circ}\text{C}$.^[41-45]

Second step: Synthesis of 4-substituted benzylidene)-6-substituted benzo[d] thiazol-2-amine

A mixture of compound (I) in step 1 (0.01M) and substituted benzaldehyde (0.02M) and 2-3 drops of glacial acid in methanol (20ml) was refluxed in a water bath for 5 hours. The solid was separated and recrystallized from ethanol.^[45-47]

Structure (plan of work)

Identification and characterization: Melting point, Solubility, thin layer chromatography, UV-spectroscopy, infrared spectroscopy, nuclear magnetic resonance spectroscopy.

Spectral Studies

Ultra Violet Spectra

Molecular absorption in the UV region of the spectrum is characteristic of the structures of the molecules. The UV scanning of the compounds was carried and 3-chloro-4-fluoroaniline exhibited λ_{max} at 265nm (Spectrum No.1).

The UV spectra of 2-amino-6-fluoro-7-chloro benzothiazole exhibited λ_{max} 303 and 288nm (Spectrum No.2). This clearly indicates the bathochromic shift of the compounds.

IR Spectra

The peaks in the IR spectrum give an idea about the

probable structure of the compound IR region ranging between 4000-666 cm^{-1} . Quanta of radiation from this region of the spectrum corresponds to energy differences between different vibrational levels of molecules. The compounds were recorded on a SHIMADZU FTIR-8400S spectrophotometer showing different vibration levels of molecules by using the KBr pellet technique. The IR spectrum of 3-chloro-4-fluoro aniline exhibited an absorption band in the region of 3433 cm^{-1} due to Ar NH₂ stretching vibrations, a band at 1622 cm^{-1} due to NH bending, banded 1494 cm^{-1} due to Ar C=C stretching, a band at 1200 due to Ar C-F stretching, a band at 762 cm^{-1} Ar C-Cl stretching (Spectrum No.3).

NMR Spectra

NMR spectroscopy enables us to record differences in magnetic properties of the various magnetic nuclei present and to deduce in a large measure the position of these nuclei are within the molecule. We can deduce how many different kinds of the environment are there in the molecules and also which atoms are present in neighboring groups. The proton NMR spectra enable us to know different chemical and magnetic environments corresponding to protons in a molecule. The samples are analyzed on Avance 300 MHz spectrometer.

Determination of partition coefficient

Procedure

Take thoroughly cleaned and well dried separating funnel and label it. Take 100ml of benzene, 100ml of distilled water, and 100mg of derivative of benzothiazole in the separating funnel. Shake the above solution mixture. Allow the solution to stand for 30 minutes for separating the

aqueous and organic layers. After 30 minutes aqueous and organic layers are separated out in different beakers. After separating out the aqueous and organic layers in different beakers label the beakers. Take 10ml of organic solution in a conical flask and titrate against 0.2N NaOH using a phenolphthalein indicator. Observe the burette readings until the color changes to pink. Note down the value of burette readings. Follow the same above procedure for an aqueous solution and note down the burette readings. After finding out the values substitute the values in the formula.^[35-37]

$$N_1V_1 = N_2V_2$$

N_1 = normality of NaOH

V_1 = Volume of NaOH Burette Readings

N_2 = Normality of organic / aqueous solution

V_2 = Volume of organic / aqueous solution

$$\text{Partition coefficient } K = C_{\text{aq}} / C_{\text{org}/1/n}$$

RESULTS AND DISCUSSION

Organic Layer

Table 1: Partition coefficient results on organic layer.

S.NO	VOLUME OF ORGANIC LAYER SOLUTION	BURETTE READINGS	AVERAGE
1.	10ml	0.4	0.4
2.	10ml	0.3	
3.	10ml	0.5	

$$N_1V_1 = N_2V_2$$

$$C_{\text{Organic}} = 1.44$$

Aqueous Layer

Table 2: Partition coefficient results on aqueous layer.

S.NO	VOLUME OF AQUEOUS LAYER	BURETTE READING	AVERAGE
1.	10ml	0.4	0.3
2.	10ml	0.2	
3.	10ml	0.3	

$$N_1V_1 = N_2V_2$$

$$C_{\text{aq}} = 1.08$$

$$\text{Partition coefficient } K = \sqrt{C_{\text{org}} / C_{\text{aq}}}$$

$$K = \sqrt{1.44 / 1.08}$$

$$K = 1.1$$

Result: As the partition coefficient value is greater than 1, the drug is more soluble in the organic phase than aqueous phase.

Table 3: Physical data compounds structure.

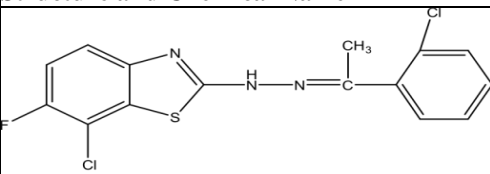
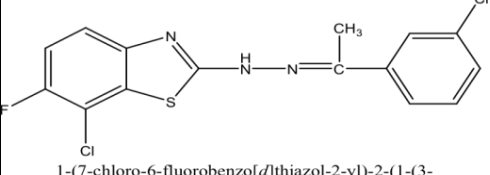
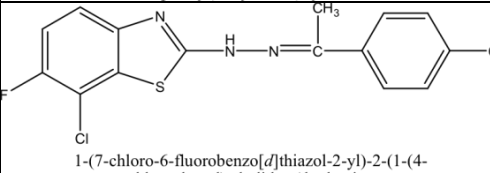
Sl. No.	Compound Code	Structure and Chemical Name
1	V D1	 1-(7-chloro-6-fluorobenzof[d]thiazol-2-yl)-2-(1-(2-chlorophenyl)ethylidene)hydrazine
2	V D2	 1-(7-chloro-6-fluorobenzof[d]thiazol-2-yl)-2-(1-(3-chlorophenyl)ethylidene)hydrazine
3	V D3	 1-(7-chloro-6-fluorobenzof[d]thiazol-2-yl)-2-(1-(4-chlorophenyl)ethylidene)hydrazine

Table 4: Solubility results.^[48]

Sl.No.	Compound Code	Soluble in	Insoluble in
1	V D1	DMF, DMSO, Ether, benzene, Acetone	Chloroform, Methanol, Ethanol
2	V D2	DMF, DMSO, Ether, benzene, Acetone	Chloroform, Methanol, Ethanol
3	V D3	DMF, DMSO, Ether, benzene, Acetone	Chloroform, Methanol, Ethanol

Table 5: Physical data of Thin layer chromatography results.

Sl.No	Compound Code	M.P/ B.P.C	% Yield	MOL. FORM	M.Wt.	C%	H%	N%
1	V D1	71-72	80%	C ₁₅ H ₁₀ S ₁ N ₃ F ₁ Cl ₂	354	50.84	2.82	11.86
2	V D2	80-82	82%	C ₁₅ H ₁₀ S ₁ N ₃ F ₁ Cl ₂	354	50.84	2.82	11.86
3	V D3	83-85	65%	C ₁₅ H ₁₀ S ₁ N ₃ F ₁ Cl ₂	354	50.84	2.82	11.86

Table 6: Analytical data results.

Sl.No	Compound Code	Solvent system for developing	Proportion of Components	Rf Value
1	V D1	Butanol Ethyl acetate Chloroform	1:2:1	0.73
2	V D2	Butanol Ethyl acetate Chloroform	1:2:1	0.76
3	V D3	Butanol Ethyl acetate Chloroform	1:2:1	0.72

Table 7: Physical data of IR spectra results.

Sl. No.	Spec. No.	Compound code	Ar- NH ₂ cm ⁻¹	ArC=C cm ⁻¹	CyclicC=N cm ⁻¹	C-F cm ⁻¹	C-Cl cm ⁻¹	NO ₂ cm ⁻¹	CH ₃ cm ⁻¹	C-N cm ⁻¹	C-O-C cm ⁻¹	Benzo-thiazole cm ⁻¹
1	03	CFA	3433	1494	-	1259	762	-	-	-	-	-
2	04	2AB	3479	1460	1646	1193	685	-	-	-	-	1390
3	05	2HB	3476	1450	1632	1194	688	-	-	-	-	1390
4	06	V D1	3350	1375	1620	1190	-	740	1175	1635	1075	1350
5	07	V D2	3425	1380	1625	1200	-	740	1175	1610	1075	1350
6	08	V D3	3360	1360	1600	1200	-	725	1175	1630	1075	1350

Table 8: NMR Spectral Data of Compounds VD₁.^[49]

Sl. No.	Spectra No.	Compound Code	Hydrogen (ppm)	Multiplicity	Solvent
1		VD1	-7H-Ar-H	6.6-7.8	Multiplet
			-1H-NH	4.2	Singlet
			-3H-CH ₃	2.85	Singlet

CONCLUSION

Synthesis characterization and lipophilicity in the present work, fluoro chloro aniline was treated with KSCN in presence of bromine in glacial acetic acid and ammonia to get 2-amino-6- fluoro-7-acid and ammonia to get 2-amino-6-fluoro-7-chloro (1,3)-benzothiazole, which was condensed with hydrazine hydrate and aldehyde in presence of con Hcl to get (4- substituted benzylidene -6-substituted benzo[d] thiazol – 2 amine). And the structure of the synthesized compounds was confirmed on the basis of spectral studies (UV, IR, NMR). And lipophilicity parameter of (4-substituted benzylidene -6-substituted benzo[d] thiazol – 2 amine) was determined by using solvents n-octanol and benzene, following the Shake flask method and titration method. Which was found that the product is distributed greater in the organic phase than in the aqueous phase, which is lipophilic in nature.

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REFERENCE

1. FILLER RJ. Fluorine Chem, 1986; 33: 361-375.
2. L. Conte, et al. J Fluorine Chem, 1995; 70: 175-179.
3. Costakes E, Tsatsas G. Chem Abstr, 1979; 90: 203935 q.
4. Mehra SC, Zaman SJ. Ind Chem Soc, 1980; 57(8): 829-32 (eng).
5. Chernykh VP, Sidorenko OF. Chem Abstr, 1983; 98: 89233.
6. Wollesdrof OW Jr, Schwam H. Chem Abstr, 1989; 111: 194656 x
7. Scholewald, Ronald D. Chem Abstr, 1984; 1002069678 c.
8. Shieke VG, Bodade AS. Chem Abstr 1991; 11423845 r.
9. Sidoova E. Chem Abstr, 1987; 106: 32907z.
10. Sidoova E. Chem Abstr, 1986; 105: 11493 c.
11. Pandey, Anil V, Lokhande SR. Indian Drugs, 1982; 19(9): 342-4 (Eng), Chem Abstr, 1983; 98: 125954.
12. Gogh T. Chem Abstr, 1981; 94: 65527.
13. Schunus, Rodney C, Gallaschun, Randall J. Chem Abstr, 191; 115: 49484 t.
14. Mouysset, Genevieve, Desagnil Sannes Gilbert., Younus Saloma, Chem Abstr 1991; 104: 122136 s.
15. Rose, Ulrich. Chem Abstr, 1991; 11529185 t.
16. Malliard, Jacques. Chem Abstr, 1985; 102: 11355n.
17. Otsuka Pharmaceutical company Ltd. Chem Abstr, 1985; 103: 196073.
18. Foscolos G, Tsatsas G. Chem Abstr, 1985; 103: 196073.
19. Oseighmah peter, Shesba, Samuel F. Chem Abstr, 1992; 116: 2356229.
20. Trivedi, Bhavani, Shah VH. Chem Abstr, 1992; 116: 151637.
21. Moharram HH. Chem Abstr, 1991; 114: 23852.
22. Chalreck P. Chem Abstr, 1988; 109: 3771.
23. Sutoris V, Halgas J. Chem Abstr, 1984; 100: 10323
24. Lipathay T, Foltinova P. Chem Abstr, 1981; 94: 10322 u.
25. Sutoris V, Foltinova P. Chem Abstr, 1981; 94: 30614
26. Sangal SK, Rastogi PK. Chem Abstr, 1980; 104: 34029 x.
27. Singh SP, Vald RK. Ind J Chem March, 1986; Vol. 25 B: PP. 228-91.
28. Greco Micheal N, Hangman William E. Chem Abstr, 1992; 117; 131109 z.
29. Sutoris V. Chem Abstr, 1991; 10683584.
30. Strelets LN, et al. Chem Abstr, 1985; 102: 78760 s.
31. Millard, Jacquece. Chem Abstr, 1985; 102: 113355 n.
32. Caryolle R, Loiseau P. Chem Abstr, 1990; 113: 237495.
33. Taked Chemical Industries Ltd. Chem Abstr, 1984; 101: 13068 p.
34. Desbois Michel, Eur Pat, APP.1 EP. 248746 (IPC Co7Co85/00) Chem Abstr, 1987; 108: 159896.
35. Sengoden Muttuswamy, et al. J Heterocyclic Chem, 1991; 28: 759.
36. Ares Jaffrey. J Chem Abstr, 1991; 114: 71453 d.
37. Shirke VG, et al. Indian Drugs, Mar 1990; 27(6).
38. Nailesh Joshi, Sushil Korgaokar and Hansa Prekh. Ind J Heterocyclic Chem, 1996; 5: 241-242.
39. Balakrishna Kalluraya, Ramesh Chimbalkar, Prashantha Gunaga. Ind J Heterocyclic Chem, 1996; 6: 103-106.
40. Dharmveer Singh, Atma Ram Mishra, Rakesh Mai Mishra. Ind J Heterocyclic Chem, 2005; 14: 289-292.
41. Mohd Afroz Bakht, Mojahidul Islam, Anees A Siddiqui. Ind J Heterocyclic Chem, 2006; 15: 297-298.
42. Bhaskar VH, Prashanth Francis, Sangameswaram B. Ind J Heterocyclic Chem, 2006; 15: 409-410.
43. Alagawadi KR, Mahajanshetti CS, Jalalpure SS. Ind J Heterocyclic Chem, 2005; 14: 315-318.
44. Dutta MM, Katakya JCS. Ind J Heterocyclic Chem, 1996; 6: 59-62.
45. Nirmala Sidker, Bulakh NR, Mehilal, Sikdar AK. Ind J Heterocyclic Chem. 2002; 12: 29-32.
46. Khan MSY, Khan RM, Sushmadrabhu. Ind J Heterocyclic Chem, 2001; 11: 119-122.
47. Shivaram Holla B, Narayana Poojary, Subramanya Bhat K. Ind J Chem, 2005; 44: 1669-1673.
48. Manjunath Bhovi, Guru S Gadaginamath. Ind J Chem, 2005; 44B: 1663-1668.
49. Jayamma Y, Sarangapani M, Reddy VM. Ind J Heterocyclic Chem, 1996; 6: 111-114.