

## PARASITIC DISEASES AND COINFECTIONS: UNRAVELING THE COMPLEX WEB OF HOST-PATHOGEN INTERACTIONS, CHALLENGES AND OPPORTUNITIES

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## ABSTRACT

Parasitic diseases and co-infections present a complex challenge to global health, requiring multidisciplinary approaches for effective control. Strengthening surveillance, improving diagnostic capabilities, and implementing integrated treatment programs are critical to mitigating their impact. This review aims to explore the epidemiology, immunological interactions, and clinical implications of parasitic diseases and co-infections. It highlights key challenges in disease management and identifies emerging opportunities for integrated approaches to prevention and treatment. A comprehensive review of peer-reviewed literature, reports from global health organizations (e.g., WHO, CDC), and recent epidemiological studies was conducted. The analysis focused on the prevalence of parasitic diseases, co-infection patterns, immunological interactions, and advancements in diagnostics and treatment strategies. Findings indicate that coinfections involving malaria, schistosomiasis, tuberculosis, HIV, and gastrointestinal parasites exacerbate disease severity and complicate treatment. The immunological interactions between parasites and other pathogens often lead to immune suppression or dysregulation, increasing susceptibility to secondary infections. Advances in molecular diagnostics, machine learning, and integrated treatment strategies are improving co-infection management. This review underscores the importance of understanding the interplay between parasitic diseases and co-infections in shaping disease outcomes. The co-occurrence of parasitic infections with other pathogens, such as bacteria, viruses, and fungi, complicates disease management, leading to increased morbidity and mortality. The intersection of these infections presents challenges in diagnosis, treatment, and public health control strategies. The findings advocate for a collaborative approach in research, healthcare policies, and public health initiatives to improve disease management and reduce the global burden of parasitic infections.

**KEYWORDS**: Parasitic Infections; Co-infections; Global Health; Molecular Diagnostics; Public Health.

## INTRODUCTION

Parasitic diseases are illnesses caused by organisms classified as parasites, which include protozoa, helminths, and ectoparasites. These organisms live on or within a host organism, deriving nutrients at the host's expense, and can cause significant morbidity and mortality worldwide. Common parasitic diseases include malaria, caused by Plasmodium species; schistosomiasis, caused by blood flukes (*Schistosoma spp.*); and intestinal parasitic infections such as those caused by *Ascaris lumbricoides* and *Giardia lamblia*. The impact of parasitic diseases is especially severe in tropical and subtropical regions, where environmental conditions, socio-economic factors, and limited access to healthcare facilitate their transmission.<sup>[1]</sup>

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On the otherhand, co-infections occur when an individual harbours two or more infectious agents simultaneously, which can lead to complex interactions between pathogens. For instance, co-infections involving parasites and other pathogens, such as viruses, bacteria, or fungi, are common in endemic areas. Examples include the coexistence of malaria and HIV, which has been shown to exacerbate immune suppression, leading to worse clinical outcomes.<sup>[2]</sup> These interactions may treatment, complicate diagnosis, and disease management, increasing the burden on healthcare systems.

The global burden of parasitic diseases remains high, with over 1 billion people affected annually.<sup>[2]</sup> Many of

these diseases overlap geographically with other infections, creating opportunities for co-infections that influence disease dynamics, treatment outcomes, and control strategies. Understanding the challenges posed by co-infections and exploring opportunities for integrated management are critical for improving global health outcomes. This review highlights key challenges in addressing parasitic diseases in the context of coinfections and identifies emerging opportunities for innovation and research.

#### 2. EPIDEMIOLOGY OF PARASITIC DISEASES AND CO-INFECTIONS

#### 2.1 Global Burden of Parasitic Diseases

Parasitic diseases significantly impact global health, with an estimated 1 billion people infected annually, predominantly in low- and middle-income countries.<sup>[2]</sup> Malaria, caused by Plasmodium species, remains a leading parasitic disease, with over 247 million cases and 619,000 deaths reported in 2021, most occurring in sub-Saharan Africa.<sup>[2]</sup> Helminth infections, such as those caused by *Ascaris lumbricoides* and *Trichuris trichiura*, affect approximately 1.5 billion people, contributing to malnutrition, anaemia, and impaired cognitive development in children.<sup>[3]</sup>

Parasitic diseases are predominantly found in regions with warm, humid climates that provide ideal conditions for the life-cycle of parasites and their vectors. The highest burden is seen in Sub-Saharan Africa, South Asia, Southeast Asia, and parts of Latin America. Each region has distinct parasitic diseases based on environmental and socioeconomic factors.

- Sub-Saharan Africa: High prevalence of malaria, schistosomiasis, lymphatic filariasis, and soil-transmitted helminths.
- South Asia: Endemic soil-transmitted helminths, leishmaniasis, and malaria.
- Southeast Asia: Malaria, schistosomiasis, and strongyloidiasis are common.
- Latin America: High prevalence of Chagas disease, leishmaniasis, and soil-transmitted helminths.

For example, malaria is endemic in over 85 countries and territories, with Sub-Saharan Africa bearing 95% of the global malaria burden.<sup>[4]</sup> Similarly, schistosomiasis is prevalent in regions where water sources are contaminated with human waste, affecting nearly 240 million people globally.<sup>[5]</sup>

Co-infections are common in endemic regions where multiple parasites overlap, further complicating the burden of these diseases. For example, individuals living in regions endemic for schistosomiasis and malaria often experience co-infections due to overlapping geographic distributions of these parasites.

The burden of parasitic diseases is closely linked to poverty, poor sanitation, and lack of access to clean water. Vulnerable populations, such as children, pregnant

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women, and immunocompromised individuals, are at higher risk of infection and severe outcomes. Key factors influencing the global burden include: i). Poverty and Malnutrition Parasitic diseases perpetuate cycles of poverty by causing chronic illness, malnutrition, and reduced productivity; ii). Lack of Access to Healthcare: Limited healthcare infrastructure in endemic regions leads to delayed diagnosis and inadequate treatment; iii). Climate Change: Rising temperatures and changing rainfall patterns affect the distribution of vectors, such as mosquitoes and snails, thereby altering the epidemiology of parasitic diseases.<sup>[6]</sup>

However, only certain population types are more susceptible to parasitic infections due to their living conditions, occupation, or health status. They include.

- i. Children: Children are particularly vulnerable to soil-transmitted helminths and malaria. Chronic parasitic infections in children can lead to stunted growth, cognitive impairment, and reduced school performance.<sup>[7]</sup>
- ii. Pregnant Women: Malaria during pregnancy increases the risk of maternal mortality, stillbirth, and low birth weight. Hookworm infections can cause iron-deficiency anemia in pregnant women, further complicating pregnancies.<sup>[8]</sup>
- iii. Immunocompromised Individuals: Individuals with compromised immune systems, such as those living with HIV/AIDS, are at higher risk of severe parasitic infections and co-infections.

#### 2.2 Common Parasitic Co-infections

Co-infections involving parasites and other pathogens are particularly prevalent in regions where environmental, socio-economic, and healthcare challenges intersect. These co-infections not only exacerbate the health burden on affected individuals but also complicate disease management and control efforts. Below are some detailed examples of common parasitic co-infections and their implications.

#### HIV and Malaria

The co-infection of HIV and malaria represents a significant public health challenge, particularly in sub-Saharan Africa where both diseases are endemic. The interaction between these two diseases exacerbates the health burden, complicates disease management, and influences both morbidity and mortality rates.<sup>[9]</sup>

HIV impairs the immune system, reducing the ability to control *Plasmodium* infections, thereby increasing the severity and frequency of malaria episodes. Conversely, acute malaria episodes can elevate HIV viral loads, likely due to immune activation and inflammatory responses triggered by the malaria infection.<sup>[10]</sup> Co-infected individuals, particularly pregnant women, are at higher risk of maternal anaemia, low birth weight in infants, and increased mortality.<sup>[11]</sup> Sub-Saharan Africa, where HIV prevalence is high and malaria transmission is endemic, is a hotspot for this co-infection.

HIV-infected individuals are at a higher risk of severe malaria infections, which can result in complications such as cerebral malaria, severe anemia, and multiorgan failure.<sup>[12]</sup> Studies have also shown that HIV-infected individuals have higher malaria parasite densities, increasing the risk of severe disease outcomes.<sup>[13]</sup>

The physiological interactions between HIV and malaria are complex and involve mutual enhancement of disease progression. HIV depletes CD4+ T cells, which play a crucial role in controlling malaria infections. This immune suppression increases susceptibility to both *Plasmodium falciparum* and *Plasmodium vivax* infections. Malaria triggers the release of proinflammatory cytokines such as TNF- $\alpha$  and IL-6, which can accelerate HIV replication.<sup>[14]</sup> This creates a feedback loop that worsens both infections. Both diseases contribute to anemia through different mechanisms. Malaria causes hemolysis and destruction of red blood cells, while HIV affects the bone marrow's ability to produce red blood cells, leading to severe anemia and chronic fatigue.<sup>[15]</sup>

The co-infection poses challenges in the treatment and control of both diseases as there are significant interactions between antiretroviral therapy (ART) and antimalarial drugs. For example, protease inhibitors used in HIV treatment can alter the metabolism of antimalarial drugs such as artemether-lumefantrine, potentially reducing efficacy.<sup>[16]</sup> Also, HIV-malaria co-infection may contribute to the emergence of drug-resistant malaria strains, particularly in regions with high ART coverage.

## Helminths and Tuberculosis (TB)

The co-infection of helminths and tuberculosis (TB) poses a significant challenge in public health, particularly in regions where both diseases are endemic. Helminth infections, which include intestinal worms such as Ascaris lumbricoides, hookworms, and Schistosoma spp., can modulate the host's immune response in ways that compromise the ability to control Mycobacterium tuberculosis infection. This interaction has important consequences for TB progression, treatment efficacy, and vaccine development. Helminth infections suppress the Th1 immune response, which is essential for controlling TB. This makes individuals more vulnerable to TB infection and progression.<sup>[17]</sup> Individuals co-infected with helminths and TB often experience slower clinical improvement during TB treatment due to immune suppression caused by helminths.<sup>[18]</sup> Chronic helminth infections can worsen TB symptoms, including persistent cough, weight loss, and fever, as helminths induce chronic inflammation and anemia.<sup>[19]</sup>

The physiological interaction between helminths and TB is primarily driven by their opposing effects on the immune system. Helminth infections induce a Type 2 helper T cell (Th2) response, which promotes the

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production of anti-inflammatory cytokines like IL-4, IL-5, and IL-10. In contrast, TB requires a Type 1 helper T cell (Th1) response, which activates macrophages and promotes the secretion of pro-inflammatory cytokines such as IFN- $\gamma$  and TNF- $\alpha$  to control the infection. Helminth infections can polarize the immune system toward a Th2 response, reducing the effectiveness of the Th1 response necessary to control TB.<sup>[19]</sup> TB infection relies on the activation of macrophages to form granulomas, which contain the bacteria. Helminthinduced IL-10 suppresses macrophage activation, reducing the host's ability to control TB.<sup>[20]</sup>

Helminth infections contribute to malnutrition, anemia, and chronic inflammation, all of which can weaken the immune system and increase the risk of TB progression. Helminths. particularly hookworms, cause iron deficiency anemia and protein-energy malnutrition, both of which impair the immune response to TB.<sup>[21]</sup> Helminth infections can cause chronic low-grade inflammation, which may accelerate the progression of latent TB to active disease.<sup>[7]</sup> The interplay between helminths and TB creates a feedback loop that worsens both infections. Helminth infections can suppress protective immunity against TB, resulting in higher bacterial loads, more severe symptoms, and longer recovery times.<sup>[19]</sup> TB infection can weaken the immune system, making the host more vulnerable to chronic helminth infections and increasing helminth-induced pathology. Helminth infections can increase the risk of latent TB progressing to active TB disease. This is particularly concerning in individuals with co-infections in endemic areas. Helminth infections can reduce the efficacy of the Bacillus Calmette-Guérin (BCG) vaccine for TB. Helminth infections impair the Th1 response required for BCG-induced immunity, resulting in a weaker protective effect.<sup>[22]</sup> Studies have shown that individuals infected with helminths in Africa and Asia have a lower immune response to BCG vaccination, indicating that helminth infections may undermine TB control efforts.[23]

## Malaria and Schistosomiasis

The co-infection of malaria and schistosomiasis is prevalent in tropical and subtropical regions, particularly in sub-Saharan Africa, where both diseases are endemic. These two parasitic diseases affect millions of people, often in overlapping geographical areas, leading to increased disease severity and complications when coinfection occurs. The interaction between Plasmodium spp. and Schistosoma spp. has significant implications for the immune system, disease progression, and treatment outcomes. The co-infection of malaria and schistosomiasis can result in more severe clinical outcomes and increased morbidity, especially in children and pregnant women Individuals co-infected with malaria and schistosomiasis experience more severe forms of both diseases, including severe anemia, organ damage, and chronic inflammation.<sup>[24]</sup> Studies show that co-infection can increase mortality rates due to

complications such as hepatic fibrosis, renal failure, and cerebral malaria.<sup>[25]</sup> Co-infection is particularly dangerous for pregnant women, increasing the risk of maternal anemia, low birth weight, preterm delivery, and infant mortality.<sup>[26]</sup> increased Malaria and schistosomiasis interact at the immune system and organ level, with each disease altering the host's physiological responses in ways that worsen the other. Malaria primarily induces a Th1 immune response, which promotes the release of pro-inflammatory cytokines like IFN- $\gamma$  and TNF- $\alpha$  to control the *Plasmodium* parasites. Schistosomiasis induces a Th2 immune response, characterized by anti-inflammatory cytokines like IL-4, IL-5, and IL-10, which help the host tolerate the presence of Schistosoma eggs but also suppress the Th1 response needed to fight malaria. This immune polarization leads to a weakened immune response against malaria parasites in individuals with schistosomiasis, resulting in higher malaria parasite loads and more severe malaria symptoms.<sup>[27]</sup> Both malaria and schistosomiasis cause hepatosplenomegaly (enlargement of the liver and spleen), which can impair immune cell function and increase the risk of organ rupture in severe cases.<sup>[28]</sup> Malaria causes red blood cell destruction and hemolysis, while schistosomiasis can cause chronic blood loss due to intestinal bleeding and urinary tract damage.<sup>[24]</sup> Coinfection can result in severe anemia, which is particularly dangerous for children and pregnant women. Malaria and schistosomiasis co-infection creates a synergistic effect that worsens the progression and outcomes of both diseases. Studies have shown that individuals with chronic schistosomiasis infections have higher malaria parasite densities and more frequent malaria episodes.<sup>[25]</sup> Schistosomiasis can impair the host's ability to clear malaria parasites from the bloodstream, leading to longer disease durations and recurrent infections.<sup>[27]</sup> Both diseases cause chronic inflammation, which can lead to immune exhaustion, making the host less capable of fighting off either infection effectively. The co-infection poses challenges for disease control and treatment. Antimalarial drugs like artemether-lumefantrine may have reduced efficacy in individuals with schistosomiasis, as liver damage caused by schistosomiasis can impair drug metabolism.<sup>[26]</sup> Mass deworming campaigns aimed at reducing schistosomiasis prevalence can also improve malaria outcomes by reducing immune suppression caused by chronic helminth infections.<sup>[27]</sup>

## Parasitic Infections and Gastrointestinal Pathogens

Parasitic infections and gastrointestinal (GI) pathogens often co-occur in tropical and developing regions, leading to significant public health challenges. The combination of intestinal parasitic infections (e.g., *Giardia lamblia, Entamoeba histolytica, Ascaris lumbricoides*) and bacterial GI pathogens (e.g., *Escherichia coli, Salmonella spp., Shigella spp.*) can result in more severe disease outcomes due to complex interactions between the pathogens and the immune system, along with the additional burden they place on

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the gastrointestinal tract. This interplay often exacerbates symptoms, complicates diagnosis and treatment, and can lead to chronic gastrointestinal disorders.<sup>[29]</sup> The coinfection of parasitic and bacterial GI pathogens can result in severe gastrointestinal symptoms, including diarrhea, malabsorption, and chronic inflammation, with increased risk of dehydration and electrolyte imbalances. Co-infected individuals often experience more severe diarrhea, prolonged symptoms, and greater malnutrition than those with only one pathogen.<sup>[30]</sup> The symptoms of co-infection can be nonspecific and overlap, making it challenging to diagnose both infections accurately. Treatment may need to address multiple pathogens simultaneously, and there can be potential drug issues.<sup>[31]</sup> interactions or resistance Also. in immunocompromised individuals, such as those with HIV/AIDS, co-infections can lead to severe dehydration, malnutrition, and chronic enteropathies, increasing the risk of mortality.<sup>[32]</sup>

The physiological effects of co-infection between parasitic and gastrointestinal pathogens are often related to immune modulation, intestinal damage, and nutrient malabsorption. Both parasitic infections and bacterial GI pathogens can alter the host's immune response, potentially weakening the body's ability to clear either infection effectively. Parasitic infections such as Giardia or Ascaris induce a Th2 immune response, which promotes anti-inflammatory cytokine production (e.g., IL-4, IL-5, IL-10) to minimize tissue damage. However, this response may dampen the host's ability to mount a sufficient immune response against bacterial pathogens.<sup>[33]</sup> In contrast, bacterial GI pathogens like Salmonella or Shigella induce a Th1 response, characterized by pro-inflammatory cytokines such as TNF- $\alpha$  and IFN- $\gamma$ , which help control bacterial growth but can exacerbate inflammation in the presence of parasitic infections.<sup>[34]</sup> The immune imbalance caused by co-infection may increase the susceptibility to severe inflammation and tissue damage, resulting in prolonged and more severe gastrointestinal symptoms.

Both types of pathogens affect the integrity of the gastrointestinal tract, leading to intestinal inflammation, intestinal permeability, and altered gut microbiota. Parasitic infections like *Giardia* and *Entamoeba histolytica* damage the intestinal epithelium, leading to malabsorption and chronic diarrhea.<sup>[35]</sup> Ascaris infections can also cause mechanical obstruction, contributing to intestinal distress. Bacterial pathogens such as *Shigella* and *Salmonella* cause intestinal inflammation and ulceration, further impairing the gut's ability to absorb nutrients and causing bleeding and watery stools.<sup>[36]</sup> Together, the combined damage from both types of pathogens can result in severe gastrointestinal dysfunction, with symptoms such as bloody diarrhea, intestinal cramping, and weight loss.

The co-infection of parasitic infections and gastrointestinal pathogens often results in synergistic

effects, leading to worsened clinical outcomes for the host. Both parasitic and bacterial infections can promote inflammatory responses that damage the intestinal mucosa. Co-infection can result in severe inflammatory responses, leading to chronic enteritis, tissue necrosis, and even perforation in extreme cases.<sup>[32]</sup> Co-infection often results in malnutrition, as the intestinal damage caused by both parasites and bacteria impairs the gut's ability to absorb nutrients. This is especially problematic in young children and immunocompromised individuals, who are more vulnerable to growth failure and developmental delays.<sup>[36]</sup> Both parasitic and bacterial infections can alter the composition of the gut microbiome, leading to dysbiosis, which impairs normal gut function and contributes to persistent gastrointestinal symptoms.<sup>[37]</sup>

Co-infection between parasitic infections and bacterial GI pathogens presents significant challenges for both disease control and treatment. Some bacterial pathogens, such as *Shigella* and *Salmonella*, are increasingly resistant to commonly used antibiotics. Co-infection with parasites can complicate treatment regimens, requiring more complex and often more expensive therapies.<sup>[30]</sup> There is a need for integrated treatment strategies that address both parasitic and bacterial infections, especially in regions with high endemicity of both types of pathogens. Current vaccines for *Shigella* and *Salmonella* are limited, and there is no vaccine for *Giardia* or *Ascaris*, highlighting a gap in preventative care.<sup>[37]</sup>

## Visceral Leishmaniasis and HIV

Visceral leishmaniasis (VL), also known as kala-azar, is a severe parasitic disease caused by Leishmania donovani and Leishmania infantum, transmitted through sandfly bites. It primarily affects the spleen, liver, and leading marrow, bone to fever. anemia, hepatosplenomegaly, and immunosuppression. The coinfection of HIV and VL is a significant global health concern, particularly in endemic regions such as East Africa, India, and the Mediterranean, where both diseases are highly prevalent.<sup>[38]</sup> Individuals with HIV/AIDS are at a higher risk of developing visceral leishmaniasis, experiencing more severe disease progression, frequent relapses, and poorer treatment outcomes.<sup>[39]</sup> Conversely, VL exacerbates HIV progression, increasing viral replication and further weakening the immune system. The interaction between VL and HIV has significant health consequences, leading to increased morbidity and mortality. HIV-infected individuals are more susceptible to Leishmania infection due to T-cell depletion (CD4+ decline), which reduces their ability to control parasitic infections.<sup>[40]</sup> Co-infected patients often exhibit disseminated leishmaniasis affecting non-typical sites such as the skin, lungs, and gastrointestinal tract, leading to misdiagnosis and delayed treatment.<sup>[41]</sup> HIV-positive individuals treated for VL have high relapse rates (60-70%), with recurrent therapy, making despite infections long-term management challenging.<sup>[38]</sup> VL contributes to higher

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viral load and faster HIV progression, increasing the risk of opportunistic infections and AIDS-related complications.<sup>[42]</sup> Co-infected patients have significantly higher mortality rates compared to those with only VL or HIV alone, with some studies reporting case-fatality rates above 50%.<sup>[39]</sup>

The co-infection of VL and HIV leads to immune system dysregulation, facilitating the progression of both diseases. VL suppresses cell-mediated immunity, reducing CD4+ T cells, which are already depleted in HIV patients, leading to severe immunosuppression.<sup>[40]</sup> Macrophages serve as reservoirs for both HIV and Leishmania, allowing both pathogens to replicate intracellularly, exacerbating immune dysfunction.<sup>[43]</sup> HIV-VL co-infection leads to a defective Th1 immune response, reducing IFN- $\gamma$  production, which is crucial for controlling Leishmania infections.<sup>[41]</sup>

VL triggers immune activation, increasing TNF- $\alpha$  levels, which enhances HIV replication and speeds up CD4+ cell decline.<sup>[42]</sup> Antigen-presenting cells (APCs), particularly dendritic cells, become dysfunctional, impairing the body's ability to mount an effective immune response against both pathogens.<sup>[43]</sup>

VL causes hepatosplenomegaly, worsening HIV-related immune dysfunction.<sup>[39]</sup> Both VL and HIV contribute to bone marrow suppression, leading to pancytopenia (anemia, leukopenia, thrombocytopenia) and increasing susceptibility to secondary infections.<sup>[38]</sup>

The bidirectional relationship between VL and HIV creates a vicious cycle that worsens clinical outcomes. HIV-positive individuals fail to mount a strong immune response against Leishmania, leading to higher parasite burden and more severe disease.<sup>[42]</sup> Standard VL treatments, such as pentavalent antimonials, are less effective in HIV patients, requiring prolonged therapy and combination regimens.<sup>[43]</sup> The combination of HIV-related immune suppression and VL-induced inflammation increases susceptibility to secondary bacterial, fungal, and viral infections, worsening morbidity.<sup>[40]</sup>

The management of HIV-VL co-infection is complicated due to the fact that HIV patients respond poorly to pentavalent antimonials and amphotericin B, requiring prolonged or higher-dose treatments.[38] Even with treatment, VL recurrence is common, necessitating lifelong secondary prophylaxis.<sup>[39]</sup> Antiretroviral therapy VL outcomes, but drug-drug (ART) improves between ART and VL medications interactions regimens.<sup>[43]</sup> complicate treatment Liposomal amphotericin B (L-AmB) is now recommended for HIV-VL co-infected patients, often combined with miltefosine for better efficacy.<sup>[41]</sup>

#### Malaria and Helminths

Helminth infections such as hookworm (*Necator americanus*) can exacerbate malaria-induced anaemia due to their additive effects on red blood cell depletion and nutrient deficiencies. Immunomodulatory effects of helminths may also alter susceptibility and severity of malaria infection.<sup>[44]</sup> Children in co-endemic regions face higher risks of severe anaemia and impaired school performance. Rural tropical regions with poor nutrition and sanitation.

#### **2.3. Environmental and Socio-economic Determinants of Parasitic Co-infections**

Environmental and socio-economic factors play pivotal roles in driving the prevalence and distribution of parasitic diseases and their co-infections. These determinants create conditions conducive to disease transmission and affect healthcare access, perpetuating a cycle of poverty and ill-health.

#### 2.3.1. Environmental Determinants

Environmental factors such as climate, geography, and infrastructure significantly influence the distribution of parasitic diseases.

#### Climate and Seasonality

Parasitic diseases like malaria and schistosomiasis thrive in tropical and subtropical regions due to favourable conditions for vectors and intermediate hosts. High humidity, warm temperatures, and seasonal rainfall create breeding grounds for mosquitoes and freshwater snails. Consequently, populations in these areas face year-round or seasonal co-infection risks. For example, in sub-Saharan Africa, co-endemicity of malaria and schistosomiasis is common due to overlapping environmental niches.<sup>[45,46]</sup>

#### Urbanization and Sanitation

Rapid urbanization without proper sanitation facilities increases the risk of soil-transmitted helminths and water-borne Protozoal infections. Poor drainage systems in urban slums contribute to stagnant water, facilitating vector breeding and outbreaks of diseases like dengue and lymphatic filariasis. Moreover, the contamination of water supplies exacerbates co-infections involving intestinal protozoa like *Giardia* and *Entamoeba*.<sup>[47]</sup>

#### **Deforestation and Agricultural Practices**

Land-use changes, such as deforestation and irrigation for agriculture, disrupt ecosystems and bring humans into closer contact with vectors and reservoirs of parasitic diseases. This has been associated with the emergence or re-emergence of zoonotic parasitic infections and co-infections, particularly in rural and peri-urban areas.<sup>[45]</sup>

#### 2.3.2. Socio-economic Determinants

Socio-economic factors often underlie vulnerability to parasitic co-infections. Few of these socio-economic determinants are discussed below.

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#### Poverty and Malnutrition

Poverty limits access to clean water, sanitation, and healthcare, all of which are crucial for preventing and managing parasitic infections. Malnutrition, prevalent in impoverished communities, exacerbates susceptibility to infections by compromising immune defences. For instance, protein-energy malnutrition has been linked to increased morbidity in helminth-TB co-infections.<sup>[48]</sup>

#### **Education and Awareness**

Low literacy rates hinder public health campaigns aimed at controlling diseases. A lack of knowledge about preventive measures, such as using insecticide-treated nets or practising proper hygiene, perpetuates the cycle of infection and co-infection.<sup>[49]</sup>

#### Healthcare Access and Inequality

Limited healthcare infrastructure in rural or marginalized areas delays diagnosis and treatment of parasitic diseases. This allows infections to progress and co-infections to become more severe. In addition, financial barriers prevent individuals from seeking treatment, further worsening health outcomes.<sup>[48]</sup>

#### 2.3.3. Behavioural and Cultural Practices

Behavioural and cultural factors also contribute to parasitic co-infections. Practices such as open defecation, bathing in contaminated water, and reliance on traditional medicine instead of biomedical treatment sustain transmission cycles. For example, cultural resistance to vaccination and drug interventions can hinder efforts to control diseases like schistosomiasis and lymphatic filariasis in endemic areas.<sup>[49]</sup>

#### 3. IMMUNOLOGICAL AND CLINICAL CHALLENGES IN PARASITIC CO-INFECTIONS 3.1. Immune System Dynamics in Co-infections

Parasitic co-infections present unique challenges to the immune system due to their ability to modulate host immunity. These interactions can either exacerbate or suppress the immune response, influencing the clinical outcomes of the diseases involved.



Figure 1: Diagramatic Illustration of parasitic coinfections and the immune response.

#### Immune Suppression by Helminths

Helminths, such as *Schistosoma* and *Ascaris*, are known to induce regulatory T cells (Tregs) and secrete immunomodulatory molecules, shifting the immune response towards a Th2-dominated profile. This can impair the host's ability to mount effective Th1 responses needed to combat intracellular pathogens, such as *Mycobacterium tuberculosis* and *Plasmodium falciparum*.<sup>[19,50]</sup> Such immune modulation is associated with increased severity of co-infections like TB and malaria.

#### Inflammatory Imbalances in Protozoal Infections

Protozoal infections, such as malaria and leishmaniasis, often induce robust inflammatory responses. When paired with helminth infections, these responses can become dysregulated, either intensifying tissue damage or allowing pathogens to evade clearance. For example, co-infection with *Plasmodium* and *Schistosoma* can result in enhanced anaemia and hepatosplenomegaly due to amplified immune activation and cytokine production.<sup>[51]</sup>

## Impact of HIV on Parasitic Co-infections

In immunocompromised individuals, particularly those with HIV/AIDS, parasitic co-infections are more severe. The depletion of CD4+ T cells compromises the host's ability to control parasitic infections, leading to higher parasite burdens and prolonged disease courses. For example, HIV accelerates the progression of visceral leishmaniasis, with co-infected individuals often experiencing higher mortality rates.<sup>[52]</sup>

# **3.2.** Clinical Manifestations, Diagnostic and Therapeutic Challenges

Parasitic co-infections alter the clinical presentation and disease trajectory, complicating diagnosis, management, and treatment. The clinical manifestations of such co-infections include.

#### a) Exacerbation of Symptoms

Co-infections can exacerbate the symptoms of individual parasitic diseases. For instance, co-infection with malaria and helminths increases the risk of severe anaemia due to additive effects on red blood cell destruction and nutrient depletion. Similarly, co-endemic schistosomiasis and tuberculosis can lead to increased granulomatous inflammation, resulting in more extensive lung damage.<sup>[44]</sup>

## b) Prolonged Disease Course and Complications

Co-infections often lead to prolonged disease courses. For example, in malaria-schistosomiasis co-infections, the presence of Schistosoma eggs in the liver exacerbates hepatic inflammation, delaying recovery from malaria. Similarly, HIV and toxoplasmosis co-infections frequently result in persistent neurological deficits due to synergistic pathogen interactions.<sup>[53]</sup>

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#### c) Increased Mortality Rates

The compounded effects of immune dysregulation and heightened symptomatology increase mortality rates in co-infected individuals. For example, visceral leishmaniasis-HIV co-infections are associated with nearly double the mortality rate of leishmaniasis alone, reflecting the synergistic impact of immune suppression.<sup>[38]</sup>

The overlapping symptomatology and altered disease progression in parasitic co-infections present significant diagnostic and therapeutic challenges including diagnositic complexity, drug-drug interactions and Resistance.

## Diagnostic Complexity

Co-infections often present with non-specific or overlapping symptoms, such as fever, anaemia, and hepatosplenomegaly, making clinical diagnosis difficult. Laboratory diagnostics are further complicated by crossreactivity in serological tests and the need for multiple, often unavailable, diagnostic tools to identify coinfecting pathogens.<sup>[54]</sup>

## Drug-Drug Interactions and Resistance

Treating co-infections requires careful consideration of drug interactions and potential toxicity. For example, anti-malarial and anti-helmintic drugs may have overlapping toxicities, requiring dose adjustments. Additionally, the rise of drug resistance in parasites like *Plasmodium falciparum* complicates treatment regimens for co-infections, necessitating more complex and costly therapeutic approaches.<sup>[55]</sup>

#### Healthcare Resource Limitations

In regions where parasitic co-infections are most prevalent, healthcare infrastructure is often inadequate. Limited access to diagnostic tools and therapeutics further exacerbates disease burden, with patients often relying on suboptimal or incomplete treatments.

#### 4. STRATEGIES FOR PREVENTION, MANAGEMENT, AND CONTROL OF PARASITIC CO-INFECTIONS

#### 4.1. Integrated Disease Control Programs

Addressing parasitic co-infections requires integrated approaches that target overlapping disease burdens. Such steps include.

## Mass Drug Administration (MDA)

Programs like MDA have shown promise in reducing the prevalence of parasitic diseases in endemic regions. For instance, distributing anti-helmintics alongside antimalarials can address co-infections in populations co-endemic for soil-transmitted helminths and malaria. However, these programs face challenges like drug resistance, logistical constraints, and incomplete coverage.<sup>[3,54]</sup>

#### **Vector Control Interventions**

Integrated vector management (IVM) addresses diseases transmitted by shared vectors. In malaria-lymphatic filariasis co-endemic areas, using long-lasting insecticidal nets (LLINs) simultaneously reduces mosquito bites and interrupts transmission of both diseases.<sup>[56]</sup> Community-driven approaches to eliminating breeding grounds further enhance these efforts.

#### Vaccination Synergies

Vaccine development programs targeting shared immune pathways can provide cross-protection. For example, research into combining vaccines for schistosomiasis and malaria is ongoing, aiming to mitigate immune suppression caused by helminth infections.<sup>[19]</sup>

In sub-Saharan Africa, integrated control programs have shown success in addressing co-endemic malaria, schistosomiasis, and helminths. Community health workers trained to distribute medications and educate on hygiene practices have reduced disease prevalence significantly. However, challenges remain in sustaining funding and combating drug resistance.<sup>[3]</sup>

#### 4.2. Strengthening Health Systems

Healthcare systems in endemic areas must be equipped to address the complex needs of co-infected individuals. Strengthening health systems involves.

#### **Enhanced Diagnostics**

Development of multiplex diagnostic tools can streamline the detection of co-infections, reducing the time and resources required. For instance, point-of-care tests capable of detecting malaria, helminths, and HIV in a single assay could significantly improve clinical outcomes.<sup>[54]</sup>

#### Capacity Building and Training

Training healthcare workers on recognizing and managing co-infections ensures appropriate and timely treatment. Emphasis on multidisciplinary teams that can handle the interplay of parasitic and non-parasitic diseases is critical.<sup>[57]</sup>

#### 4.3. Research and Policy Innovations

Ongoing research and innovative policies are essential to tackle the evolving challenges of parasitic co-infections.

#### a) Operational Research

Studies focusing on co-infection dynamics help identify high-risk populations and optimize resource allocation. For example, mapping co-endemic regions aids in deploying integrated interventions effectively.<sup>[58]</sup>

#### b) Global Policy Frameworks

Policies such as the WHO's Roadmap for Neglected Tropical Diseases emphasize integrating parasitic disease control into broader health agendas, fostering collaboration across sectors.<sup>[54]</sup>

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#### 5: OPPORTUNITIES IN RESEARCH AND INNOVATION FOR MANAGING PARASITIC CO-INFECTIONS

## 5.1. Advancements in Diagnostics, Molecular and Genomic Tools

Recent innovations in molecular and genomic tools have revolutionized the study and management of parasitic diseases, offering unprecedented opportunities to understand co-infection dynamics.

#### a) Next-Generation Sequencing (NGS)

NGS technologies allow detailed profiling of parasitic communities and their interactions with hosts. Metagenomic studies can identify co-infections by detecting multiple pathogen genomes from a single improving diagnostic sample. precision and surveillance.<sup>[59]</sup> These tools have uncovered novel associations between parasitic species and their impact on disease outcomes. For example, genome-wide association studies (GWAS) have revealed host genetic factors that influence susceptibility to co-infections, such as malaria and helminths, paving the way for personalized interventions.<sup>[60]</sup>

#### b) CRISPR-Cas9 in Parasitology

Gene-editing tools like CRISPR-Cas9 are being applied to study gene function in parasites, such as *Plasmodium falciparum* and *Leishmania*. These approaches can help identify virulence factors and drug resistance genes, facilitating the development of targeted therapies.<sup>[61]</sup> Additionally, CRISPR technology is being explored to create gene-drive systems for vector control, aiming to reduce the transmission of parasitic diseases.<sup>[62]</sup>

## c) Multiplex Diagnostic Platforms

Multiplex PCR platforms, such as the xTAG system, have been used successfully to detect co-infections by amplifying multiple DNA targets in a single reaction. For instance, this method was shown to simultaneously detect *Plasmodium falciparum* and intestinal protozoa in co-endemic regions.<sup>[65]</sup> Similarly, multiplex ELISA tests now combine malaria-specific antigens and helminth markers, improving diagnostic throughput for clinics in sub-Saharan Africa.<sup>[66]</sup>

#### d) Point-of-Care Devices

Advances in portable diagnostics, including CRISPRbased systems, enable real-time and cost-effective identification of co-infections. For example, a CRISPRpowered lateral flow assay developed in 2021 successfully detected malaria and schistosomiasis antigens within an hour, providing a promising tool for field applications.<sup>[67]</sup>

#### e) Biomarker Identification

Studies in proteomics have identified glycoproteins unique to co-infections, such as the combined malariaschistosomiasis biomarker, aiding in developing pointof-care multiplex kits. Such advancements reduce diagnostic errors and ensure timely intervention.<sup>[25]</sup>

## **5.2. Development of Integrated Treatment and Multi-Target Therapeutics**

Co-infections often require simultaneous treatment of multiple pathogens. The development of multi-target therapeutics offers a promising solution to this challenge.

## a) Combination Therapies

Studies report enhanced efficacy when combining artemisinin-based antimalarials with praziquantel for treating malaria-schistosomiasis co-infections, leading to significant reductions in parasitic load and faster recovery.<sup>[68]</sup> Additionally, the integration of ivermectin for filariasis and malaria has shown success in joint elimination programs.<sup>[69]</sup>

## b) Broad-Spectrum Anti-parasitic Drugs

Efforts to develop drugs that target conserved pathways across parasitic species are gaining momentum. For instance, inhibitors of glycolytic enzymes, essential for energy metabolism in both protozoa and helminths, are being investigated as potential broad-spectrum agents.<sup>[63]</sup>

## c) Nanotechnology in Drug Delivery

Nanotechnology offers innovative solutions for drug delivery in parasitic diseases. Nanocarriers can enhance the bioavailability and targeted delivery of anti-parasitic drugs, reducing side effects and improving efficacy. For example, liposomal formulations of amphotericin B are used to treat visceral leishmaniasis with reduced toxicity.<sup>[64]</sup>

## d) Pharmacogenomics

A novel approach leverages genomic insights to personalize drug regimens for patients, especially in managing resistance-prone co-infections. For example, recent work on CYP450 enzymes has refined antimalarial and Anti-helminthic dosing strategies.<sup>[70]</sup>

# 5.3. Harnessing Big Data and Artificial Intelligence (AI)

The integration of big data and AI in parasitology is creating new avenues for understanding and managing co-infections.

#### a) Machine Learning in Diagnostics

Machine learning models trained on genomic and proteomic datasets are improving diagnostic accuracy for co-infections. For example, AI-driven image analysis of blood smears has been used to differentiate between Plasmodium species and detect co-infecting helminths.<sup>[71]</sup>

## b) Big Data Analytics

Platforms integrating climatic, demographic, and epidemiological data provide predictive analytics for hotspots. For example, a study using machine learning predicted malaria-lymphatic filariasis overlap in South-east Asia, informing vector control.<sup>[66]</sup>

# c) Artificial Intelligence (AI) and Machine Learning (ML)

Tools like Google's TensorFlow have been applied to

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analyze spatial co-infection patterns. In Africa, machine learning algorithms successfully mapped soil-transmitted helminths alongside malaria incidence based on satellite data and environmental variables.<sup>[70]</sup>

## d) Remote Sensing and GIS

Remote sensing technologies, particularly Sentinel-2 satellite imaging, have proven critical in monitoring environmental changes linked to co-infection risks. For example, snail habitat mapping for schistosomiasis combined with malaria hotspot data enhanced intervention targeting in Nigeria.<sup>[60]</sup>

## **5.4.** Opportunities in Vaccine Development

Vaccine research offers significant opportunities to mitigate the burden of parasitic co-infections.

## a) Multi-Valent Vaccines

Developing multi-valent vaccines that target multiple parasites is a promising area of research. For example, efforts to combine antigens from Plasmodium and Schistosoma aim to protect against both malaria and schistosomiasis.<sup>[72]</sup>

## b) Adjuvant Strategies

Advances in adjuvant technologies are enhancing the efficacy of vaccines for parasitic diseases. Adjuvants that modulate immune responses, such as TLR agonists, are being explored to overcome the immunosuppressive effects of co-infections.<sup>[73]</sup>

## 5.5. Community-Based Approaches

#### a) Community Engagement in Surveillance

Programs in Uganda train local health workers in disease tracking via mobile applications like DHIS2. These efforts have improved real-time reporting of malaria and helminth infections, increasing the efficiency of intervention deployment.<sup>[69]</sup>

#### b) Participatory Research Models

Engaging communities in intervention planning ensures cultural acceptance and sustainability. For instance, in co-endemic regions of Nigeria, researchers collaborated with locals to develop snail control initiatives, reducing schistosomiasis prevalence by 40% in three years.<sup>[60]</sup>

#### 5.6. Strengthening Collaborative Research Networks

Collaboration among researchers, governments, and international organizations is crucial for advancing knowledge and control of parasitic co-infections.

#### a) Global Initiatives

Initiatives like the Global Alliance for Vaccines and Immunization (GAVI) and the Coalition for Epidemic Preparedness Innovations (CEPI) support research and development for neglected tropical diseases, including parasitic co-infections.

#### b) South-South Collaborations

Strengthening collaborations among endemic countries facilitates the exchange of knowledge and resources.

Regional research consortia, such as the African Network for Neglected Tropical Diseases, are instrumental in addressing the unique challenges of parasitic co-infections in resource-limited settings.<sup>[74]</sup>

# 6. CASE STUDIES AND REGIONAL PERSPECTIVES

#### 6.1 Examples of Successful Interventions

Uganda: Schistosomiasis Control Through Integrated Programs

Uganda has implemented mass drug administration (MDA) with praziquantel alongside water, sanitation, and hygiene (WASH) interventions. These programs have been complemented by community health education campaigns, addressing gender-specific risks of infection. This integrated approach has proven effective in reducing schistosomiasis reinfection rates.

Outcome: Studies emphasize that combining MDA with WASH and health promotion significantly improves disease outcomes and helps move toward elimination in highly endemic areas.<sup>[75,76]</sup>

# Cambodia: Malaria-Elimination Efforts in the Mekong Region.

In Cambodia, the national malaria control program has focused on multidrug-resistant malaria by using artemisinin-based combination therapies (ACTs) alongside enhanced surveillance and rapid diagnostic tools. Efforts concentrate on forested and remote areas where malaria prevalence is high.

Outcome: Declines in malaria incidence demonstrate the success of targeted approaches in these challenging settings.<sup>[77]</sup>

## Brazil: Lymphatic Filariasis Elimination in Recife

Brazil's program combines vector control, mass drug administration, and public awareness initiatives in urban areas like Recife. These interventions focus on combating mosquito-borne parasitic infections, particularly lymphatic filariasis.

Outcome: Near elimination of lymphatic filariasis highlights the success of sustained, integrated interventions.<sup>[78]</sup>

## 6.2 Regional Differences in Challenges and Solutions *Sub-Saharan Africa*

Challenges: High burden of malaria, schistosomiasis, and soil-transmitted helminths compounded by healthcare access issues.

Solutions: Predictive modelling helps identify high-risk areas for targeted interventions. Studies on spatial epidemiology suggest integrating big data with field-based observations to map disease hotspots effectively.<sup>[79, 80]</sup>

#### South-east Asia

Challenges: Diverse ecological settings and emergence of drug-resistant parasites complicate control measures. Solutions: Strengthened regional collaborations, such as through the Asia Pacific Malaria Elimination Network,

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and innovative tools like larvivorous fish for vector control in rice paddies are effective strategies.

#### South America

Challenges: Geographic barriers in the Amazon and coinfections with viral diseases such as dengue.

Solutions: Community-led vector management initiatives, coupled with technology-based solutions like drones for medical supply delivery, have shown promising results in overcoming logistical barriers.

#### South Asia

Challenges: Dense population centers contribute to the rapid spread of parasitic diseases.

Solutions: Implementation of portable diagnostic kits for real-time surveillance and targeted awareness campaigns address these challenges effectively.

# 7. FUTURE DIRECTIONS AND RESEARCH NEEDS

7.1 Research Gaps

## a) Understudied Interactions Between Parasites and Other Pathogens

Parasitic co-infections often interact with other infectious agents like bacteria, viruses, or fungi, leading to complex immune responses. For instance, interactions between helminths and Mycobacterium tuberculosis influence immune modulation, affecting TB progression and vaccine efficacy.<sup>[81]</sup> Additionally, parasitic infections such as schistosomiasis alter immune profiles, which may increase susceptibility to HIV acquisition or progression.<sup>[51]</sup> Understanding these immune interactions is critical to improving therapeutic approaches.

## b) Limited Insights into Chronic Co-Infection Effects

Chronic co-infections may contribute to long-term health impacts, including impaired growth in children, exacerbated anaemia, and increased susceptibility to metabolic and cardiovascular diseases.<sup>[50]</sup> Research into how parasitic infections alter host metabolism, gut microbiota, or epigenetic profiles remains sparse.

#### c) Diagnostic Challenges in Co-Infections

Current diagnostic methods often focus on a single pathogen. There is a significant gap in multiplex diagnostic platforms capable of detecting multiple parasitic pathogens and their co-infecting agents efficiently in low-resource settings.<sup>[82]</sup>

#### 7.2 Policy Recommendations

#### a) Integrated Healthcare Policies for Co-Infections

Integrated healthcare programs must prioritize coendemic areas where parasitic infections coexist with other diseases like TB or malaria. Coordinating MDA campaigns, vector control, and vaccination drives can maximize health outcomes.

Example: The WHO has advocated for integrating TB and HIV management in endemic regions, emphasizing co-morbidity reduction.<sup>[54]</sup>

## b) Global Surveillance and Resource Allocation

Policymakers should strengthen disease surveillance using advanced technologies like GIS, remote sensing, and AI-driven models. Enhanced data collection enables resource allocation to hotspots with the highest coinfection burden.

#### c) Sustainable Funding Mechanisms

Long-term commitments from global health bodies, governments, and private stakeholders are critical to sustain parasitic disease elimination programs and develop novel therapeutics.<sup>[83,84]</sup>

## 7.3 Areas for Further Investigation

## a) Genomic Studies on Host-Parasite Dynamics

Advances in genomics and transcriptomics can uncover host-pathogen interactions during co-infections. For instance, genetic variants influencing immune pathways may predispose individuals to severe outcomes.<sup>[60]</sup>

#### b) Environmental and Climate Factors

Studies must explore how environmental changes, including deforestation and urbanization, alter the spatial distribution of parasitic diseases and their co-infections.,<sup>[80, 85]</sup>

#### c) Behavioural Research in Endemic Communities

Cultural practices, traditional medicine usage, and health-seeking behaviours can impact the uptake of interventions. Behavioural studies can inform culturally sensitive and effective strategies for disease management.

## 8. CONCLUSION

The management of parasitic diseases and co-infections is hampered by diagnostic gaps, limited integrated healthcare policies, and the long-term impacts of untreated infections. Technological advancements in diagnostics, AI-driven surveillance, and community engagement provide pathways for addressing parasitic diseases. Integrating interventions with broader healthcare strategies can enhance outcomes.

A collaborative, interdisciplinary approach is essential. Policymakers, researchers, healthcare providers, and communities must work together to address research gaps, implement effective policies, and develop innovative tools for managing parasitic diseases and coinfections globally.

## REFERENCES

- Centers for Disease Control and Prevention (CDC). (2023). Parasites - Diseases. Retrieved from https://www.cdc.gov/parasites/
- World Health Organization (WHO). (2021). Malaria and HIV/AIDS Interactions and Implications. Retrieved from https://www.who.int

I

- Hotez, P. J., Fenwick, A., Savioli, L., & Molyneux, D. H. (2009). Rescuing the bottom billion through control of neglected tropical diseases. *Lancet* (*London, England*), 373(9674): 1570–1575. https://doi.org/10.1016/S0140-6736(09)60233-6
- 4. World Health Organization (WHO). (2023). Malaria. Retrieved from https://www.who.int/newsroom/fact-sheets/detail/malaria
- 5. Centers for Disease Control and Prevention (CDC). (2022). Schistosomiasis. Retrieved from https://www.cdc.gov/parasites/schistosomiasis
- Haines, A., Kovats, R. S., Campbell-Lendrum, D., & Corvalan, C. (2006). Climate change and human health: impacts, vulnerability, and mitigation. *Lancet* (*London, England*), 367(9528): 2101–2109. https://doi.org/10.1016/S0140-6736(06)68933-2
- Bethony, J., Brooker, S., Albonico, M., Geiger, S. M., Loukas, A., Diemert, D., & Hotez, P. J. (2006). Soil-transmitted helminth infections: ascariasis, trichuriasis, and hookworm. *Lancet* (London, England), 367(9521): 1521–1532. https://doi.org/10.1016/S0140-6736(06)68653-4
- Brooker, S., Hotez, P. J., & Bundy, D. A. (2008). Hookworm-related anaemia among pregnant women: a systematic review. *PLoS neglected tropical diseases*, 2(9): e291. https://doi.org/10.1371/journal.pntd.0000291
- Kwenti, T. E. (2018). Malaria and HIV coinfection in sub-Saharan Africa: prevalence, impact, and treatment strategies. *Research and reports in tropical medicine*, 9: 123–136. https://doi.org/10.2147/RRTM.S154501
- Van Geertruyden, J. P. (2014). Interactions between malaria and human immunodeficiency virus anno 2014. Clinical microbiology and infection: the official publication of the European Society of Clinical Microbiology and Infectious Diseases, 20(4): 278–285. https://doi.org/10.1111/1469-0691.12597
- Alemu, A., Shiferaw, Y., Addis, Z., Mathewos, B., & Birhan, W. (2013). Effect of malaria on HIV/AIDS transmission and progression. *Parasites* & vectors, 6: 18. https://doi.org/10.1186/1756-3305-6-18
- 12. Hochman, S., & Kim, K. (2012). The Impact of HIV Coinfection on Cerebral Malaria Pathogenesis. *Journal of neuroparasitology*, 3: 235547. https://doi.org/10.4303/jnp/235547
- Kublin, J. G., Patnaik, P., Jere, C. S., Miller, W. C., Hoffman, I. F., Chimbiya, N., Pendame, R., Taylor, T. E., & Molyneux, M. E. (2005). Effect of Plasmodium falciparum malaria on concentration of HIV-1-RNA in the blood of adults in rural Malawi: a prospective cohort study. *Lancet (London, England)*, 365(9455): 233–240. https://doi.org/10.1016/S0140-6736(05)17743-5.
- Wilairatana, P., Mala, W., Milanez, G. J., Masangkay, F. R., Kotepui, K. U., & Kotepui, M. (2022). Increased interleukin-6 levels associated with malaria infection and disease severity: a

systematic review and meta-analysis. *Scientific reports*, 12(1): 5982. https://doi.org/10.1038/s41598-022-09848-9.

- Hochman, S., & Kim, K. (2009). The Impact of HIV and Malaria Coinfection: What Is Known and Suggested Venues for Further Study. *Interdisciplinary perspectives on infectious diseases*, 2009; 617954.
- 16. Byakika-Kibwika, P., Lamorde, M., Mayito, J., Nabukeera, L., Namakula, R., Mayanja-Kizza, H., Katabira, E., Ntale, M., Pakker, N., Ryan, M., Hanpithakpong, W., Tarning, J., Lindegardh, N., de Vries, P. J., Khoo, S., Back, D., & Merry, C. (2012). Significant pharmacokinetic interactions between artemether/lumefantrine and efavirenz or nevirapine in HIV-infected Ugandan adults. *The Journal of antimicrobial chemotherapy*, 67(9): 2213–2221. https://doi.org/10.1093/jac/dks207.
- 17. Elias, D., Wolday, D., Akuffo, H., Petros, B., Bronner, U., & Britton, S. (2001). Effect of deworming on human T cell responses to mycobacterial antigens in helminth-exposed individuals before and after bacille Calmette-Guérin (BCG) vaccination. Clinical and experimental immunology, 123(2): 219-225. https://doi.org/10.1046/j.1365-2249.2001.01446.x Cadmus, S. I., Akinseye, V. O., Taiwo, B. O., Pinelli, E. O., van Soolingen, D., & Rhodes, S. G. (2020). Interactions between helminths and tuberculosis infections: Implications for tuberculosis diagnosis and vaccination in Africa. PLoS neglected tropical diseases, 14(6): e0008069. https://doi.org/10.1371/journal.pntd.0008069.
- Babu, S., & Nutman, T. B. (2016). Helminth-Tuberculosis Co-infection: An Immunologic Perspective. *Trends in immunology*, 37(9): 597–607. https://doi.org/10.1016/j.it.2016.07.005
- George, P. J., Anuradha, R., Kumar, N. P., Sridhar, R., Banurekha, V. V., Nutman, T. B., & Babu, S. (2014). Helminth infections coincident with active pulmonary tuberculosis inhibit mono- and multifunctional CD4+ and CD8+ T cell responses in a process dependent on IL-10. *PLoS pathogens*, 10(9): e1004375. https://doi.org/10.1371/journal.ppat.1004375.
- Jordao, L., Lengeling, A., Bordat, Y., Boudou, F., Gicquel, B., Neyrolles, O., Becker, P. D., Guzman, C. A., Griffiths, G., & Anes, E. (2008). Effects of omega-3 and -6 fatty acids on *Mycobacterium tuberculosis* in macrophages and in mice. *Microbes and infection*, 10(12-13): 1379–1386. https://doi.org/10.1016/j.micinf.2008.08.004.
- Elias, D., Britton, S., Aseffa, A., Engers, H., & Akuffo, H. (2008). Poor immunogenicity of BCG in helminth infected population is associated with increased in vitro TGF-beta production. *Vaccine*, 26(31): 3897–3902. https://doi.org/10.1016/j.vaccine.2008.04.083.

I

- Randall, A. E., Perez, M. A., Floyd, S., Black, G. F., Crampin, A. C., Ngwira, B., Pistoni, W. N., Mulawa, D., Sichali, L., Mwaungulu, L., Bickle, Q., & Fine, P. E. (2002). Patterns of helminth infection and relationship to BCG vaccination in Karonga District, northern Malawi. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 96(1): 29–33. https://doi.org/10.1016/s0035-9203(02)90229-4.
- 23. Kinung'hi, S. M., Mazigo, H. D., Dunne, D. W., Kepha, S., Kaatano, G., Kishamawe, C., Ndokeji, S., Angelo, T., & Nuwaha, F. (2017). Coinfection of intestinal schistosomiasis and malaria and association with haemoglobin levels and nutritional status in school children in Mara region, Northwestern Tanzania: а cross-sectional exploratory study. BMC research notes, 10(1): 583. https://doi.org/10.1186/s13104-017-2904-2
- Dassah, S. D., Nyaah, K. E., Senoo, D. K. J., Ziem, J. B., Aniweh, Y., Amenga-Etego, L., Awandare, G. A., & Abugri, J. (2023). Co-infection of *Plasmodium falciparum* and *Schistosoma mansoni* is associated with anaemia. *Malaria journal*, 22(1): 272. https://doi.org/10.1186/s12936-023-04709-w
- Friedman, J. F., Mital, P., Kanzaria, H. K., Olds, G. R., & Kurtis, J. D. (2007). Schistosomiasis and pregnancy. Trends in parasitology, 23(4): 159–164. https://doi.org/10.1016/j.pt.2007.02.006
- 26. Ateba Ngoa, U., Zinsou, J. F., Kassa, R. F., Ngoune Feugap, E., Honkpehedji, Y. J., Massinga-Loembe, M., Kenguele Moundounga, H., Nkoma Mouima, A. M., Mbenkep, L. H., Wammes, L. J., Mbow, M., Kruize, Y., Mombo-Ngoma, G., Bouyoukou Hounkpatin, A. L., Dejon Agobe, J. C., Saadou, I., Lell, B., Smits, H., Kremsner, P. G., Yazdanbakhsh, M., ... Adegnika, A. A. (2014). Assessment of the effect of Schistosoma haematobium co infection on malaria parasites and immune responses in rural populations Gabon: study in 388. protocol. SpringerPlus, 3: https://doi.org/10.1186/2193-1801-3-388
- 27. Wilson, S., Jones, F. M., Mwatha, J. K., Kimani, G., Booth, M., Kariuki, H. C., Vennervald, B. J., Ouma, J. H., Muchiri, E., & Dunne, D. W. (2009). Hepatosplenomegaly associated with chronic malaria exposure: evidence for a pro-inflammatory mechanism exacerbated by schistosomiasis. *Parasite immunology*, 31(2): 64–71. https://doi.org/10.1111/j.1365-3024.2008.01078.x
- Elmonir, W., Elaadli, H., Amer, A., El-Sharkawy, H., Bessat, M., Mahmoud, S. F., Atta, M. S., & El-Tras, W. F. (2021). Prevalence of intestinal parasitic infections and their associated risk factors among preschool and school children in Egypt. *PloS one*, 16(9): e0258037. https://doi.org/10.1371/journal.pone.0258037
- Di Genova, B. M., & Tonelli, R. R. (2016). Infection Strategies of Intestinal Parasite Pathogens and Host Cell Responses. *Frontiers in microbiology*, 7: 256. https://doi.org/10.3389/fmicb.2016.00256

- Braseth, A. L., Elliott, D. E., & Ince, M. N. (2021). Parasitic Infections of the Gastrointestinal Track and Liver. *Gastroenterology clinics of North America*, 50(2): 361–381. https://doi.org/10.1016/j.gtc.2021.02.011
- 31. Duggal, S., Chugh, T. D., & Duggal, A. K. (2012). HIV and malnutrition: effects on immune system. *Clinical & developmental immunology*, 2012; 784740. https://doi.org/10.1155/2012/784740
- Hotez, P. J., Brindley, P. J., Bethony, J. M., King, C. H., Pearce, E. J., & Jacobson, J. (2008). Helminth infections: the great neglected tropical diseases. *The Journal of clinical investigation*, 118(4): 1311–1321. https://doi.org/10.1172/JCI34261
- 33. Zhao, M., Chu, J., Feng, S., Guo, C., Xue, B., He, K., & Li, L. (2023). Immunological mechanisms of inflammatory diseases caused by gut microbiota dysbiosis: A review. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*, 164: 114985. https://doi.org/10.1016/j.biopha.2023.114985
- 34. Gutiérrez, L., & Bartelt, L. (2024). Current Understanding of Giardia lamblia and Pathogenesis of Stunting and Cognitive Deficits in Children from Low- and Middle-Income Countries. Current tropical medicine reports, 11(1): 28–39. https://doi.org/10.1007/s40475-024-00314-2
- 35. Fauziah, N., Aviani, J. K., Agrianfanny, Y. N., & Fatimah, S. N. (2022). Intestinal Parasitic Infection and Nutritional Status in Children under Five Years Old: A Systematic Review. *Tropical medicine and infectious* disease, 7(11): 371. https://doi.org/10.3390/tropicalmed7110371
- 36. Gorkiewicz, G., & Moschen, A. (2018). Gut microbiome: a new player in gastrointestinal disease. Virchows Archiv : an international journal of pathology, 472(1): 159–172. https://doi.org/10.1007/s00428-017-2277-x
- Alvar, J., Aparicio, P., Aseffa, A., Den Boer, M., Cañavate, C., Dedet, J. P., Gradoni, L., Ter Horst, R., López-Vélez, R., & Moreno, J. (2008). The relationship between leishmaniasis and AIDS: the second 10 years. *Clinical microbiology reviews*, 21(2): 334–359. https://doi.org/10.1128/CMR.00061-07
- Machado, C. A. L., Sevá, A. D. P., Silva, A. A. F. A. E., & Horta, M. C. (2021). Epidemiological profile and lethality of visceral leishmaniasis/human immunodeficiency virus co-infection in an endemic area in Northeast Brazil. *Revista da Sociedade Brasileira de Medicina Tropical*, 54: e0795. https://doi.org/10.1590/0037-8682-0795-2020
- Takele, Y., Mulaw, T., Adem, E., Womersley, R., Kaforou, M., Franssen, S. U., Levin, M., Taylor, G. P., Müller, I., Cotton, J. A., & Kropf, P. (2022). Recurrent visceral leishmaniasis relapses in HIV coinfected patients are characterized by less efficient immune responses and higher parasite

I

load. *iScience*, 26(2): https://doi.org/10.1016/j.isci.2022.105867

40. Okwor, I., & Uzonna, J. E. (2013). The immunology of Leishmania/HIV co-infection. *Immunologic research*, 56(1): 163–171. https://doi.org/10.1007/s12026-013-8389-8

105867.

- Charoensakulchai, S., Bualert, L., Manomat, J., Mungthin, M., Leelayoova, S., Tan-Ariya, P., Siripattanapipong, S., Naaglor, T., & Piyaraj, P. (2020). Risk Factors of Leishmania Infection among HIV-Infected Patients in Trang Province, Southern Thailand: A Study on Three Prevalent Species. *The American journal of tropical medicine and hygiene*, 103(4): 1502–1509. https://doi.org/10.4269/aitmh.20-0332
- Murray, H. W., Berman, J. D., Davies, C. R., & Saravia, N. G. (2005). Advances in leishmaniasis. *Lancet (London, England)*, 366(9496): 1561–1577. https://doi.org/10.1016/S0140-6736(05)67629-5
- 43. Nacher, M. (2004). Interactions between worm infections and malaria. *Clinical reviews in allergy & immunology*, 26(2): 85–92. https://doi.org/10.1007/s12016-004-0003-3
- 44. Hotez, P. J., & Kamath, A. (2009). Neglected tropical diseases in sub-saharan Africa: review of their prevalence, distribution, and disease burden. *PLoS neglected tropical diseases*, 3(8): e412. https://doi.org/10.1371/journal.pntd.0000412
- 45. Ekpunobi, N., Akinsuyi, O., Ariri, T., & Ogunmola, T. (2023). The Reemergence of Monkeypox in Nigeria. *Challenges*, 14(2): 22. https://doi.org/10.3390/challe14020022
- 46. Archer, J., O'Halloran, L., Al-Shehri, H., Summers, S., Bhattacharyya, T., Kabaterine, N. B., Atuhaire, A., Adriko, M., Arianaitwe, M., Stewart, M., LaCourse, E. J., Webster, B. L., Bustinduy, A. L., & Stothard, J. R. (2020). Intestinal Schistosomiasis and Giardiasis Co-Infection in Sub-Saharan Africa: Can a One Health Approach Improve Control of Each Waterborne Parasite Simultaneously?. *Tropical medicine and infectious disease*, 5(3): 137. https://doi.org/10.3390/tropicalmed5030137
- Lönnroth, K., & Raviglione, M. (2008). Global epidemiology of tuberculosis: prospects for control. Seminars in respiratory and critical care medicine, 29(5): 481–491. https://doi.org/10.1055/s-0028-1085700
- Chitsulo, L., Engels, D., Montresor, A., & Savioli, L. (2000). The global status of schistosomiasis and its control. *Acta tropica*, 77(1): 41–51. https://doi.org/10.1016/s0001-706x(00)00122-4.
- 49. Maizels, R. M., & McSorley, H. J. (2016). Regulation of the host immune system by helminth parasites. *The Journal of allergy and clinical immunology*, *138*(3): 666–675. https://doi.org/10.1016/j.jaci.2016.07.007
- 50. Van Riet, E., Hartgers, F. C., & Yazdanbakhsh, M. (2007). Chronic helminth infections induce immunomodulation: consequences and

mechanisms. *Immunobiology*, *212*(6): 475–490. https://doi.org/10.1016/j.imbio.2007.03.009

- 51. Diro, E., Lynen, L., Ritmeijer, K., Boelaert, M., Hailu, A., & van Griensven, J. (2014). Visceral Leishmaniasis and HIV coinfection in East Africa. *PLoS neglected tropical diseases*, 8(6): e2869. https://doi.org/10.1371/journal.pntd.0002869
- Price R. W. (1996). Neurological complications of HIV infection. *Lancet* (London, England): 348(9025): 445–452. https://doi.org/10.1016/S0140-6736(95)11035-6
- 53. WHO (2021). Integrated Diagnostics for Parasitic Co-Infections. World Health Organization.
- Murray, C. J., Vos, T., Lozano, R., Naghavi, M., Flaxman, A. D., Michaud, C., Ezzati, M., Shibuya, K., Salomon, J. A., Abdalla, S., Aboyans, V., Abraham, J., Ackerman, I., Aggarwal, R., Ahn, S. Y., Ali, M. K., Alvarado, M., Anderson, H. R., Anderson, L. M., Andrews, K. G., ... Memish, Z. A. (2012). Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet (London, England)*, 380(9859): 2197–2223. https://doi.org/10.1016/S0140-6736(12)61689-4
- 55. Sunmonu, A. B., & Ekpunobi, N. F. (2023). Larvicidal potential of silver nanoparticles synthesized from ocimum gratissimum leaf extracts against anopheles' mosquito. *GSC Biological and Pharmaceutical Sciences*, 25(3): 041-048. https://doi.org/10.30574/gscbps.2023.25.3.0517
- Liese, B., Rosenberg, M., & Schratz, A. (2010). Programmes, partnerships, and governance for elimination and control of neglected tropical diseases. *Lancet* (*London, England*): 375(9708): 67–76. https://doi.org/10.1016/S0140-6736(09)61749-9
- 57. Brooker, S., & Utzinger, J. (2007). Integrated disease mapping in a polyparasitic world. *Geospatial health*, *1*(2): 141–146. https://doi.org/10.4081/gh.2007.262
- Pallen, M. J. (2014). Diagnostic metagenomics: potential applications to bacterial, viral and parasitic infections. *Parasitology*, 141(14): 1856–1862. https://doi.org/10.1017/S0031182014000134
- 59. Jallow, M., Teo, Y. Y., Small, K. S., Rockett, K. A., Deloukas, P., Clark, T. G., Kivinen, K., Bojang, K. A., Conway, D. J., Pinder, M., Sirugo, G., Sisay-Joof, F., Usen, S., Auburn, S., Bumpstead, S. J., Campino, S., Coffey, A., Dunham, A., Fry, A. E., Green, A., ... Malaria Genomic Epidemiology Network (2009). Genome-wide and fine-resolution association analysis of malaria in West Africa. Nature genetics, 41(6): 657-665. https://doi.org/10.1038/ng.388
- Ghorbal, M., Gorman, M., Macpherson, C. R., Martins, R. M., Scherf, A., & Lopez-Rubio, J. J. (2014). Genome editing in the human malaria parasite *Plasmodium falciparum* using the CRISPR-

I

Cas9 system. *Nature biotechnology*, *32*(8): 819–821. https://doi.org/10.1038/nbt.2925

- Kyrou, K., Hammond, A. M., Galizi, R., Kranjc, N., Burt, A., Beaghton, A. K., Nolan, T., & Crisanti, A. (2018). A CRISPR-Cas9 gene drive targeting doublesex causes complete population suppression in caged Anopheles gambiae mosquitoes. *Nature biotechnology*, *36*(11): 1062–1066. https://doi.org/10.1038/nbt.4245
- 62. Springer, A. L., Agrawal, S., & Chang, E. P. (2024). Malate dehydrogenase in parasitic protozoans: roles in metabolism and potential therapeutic applications. *Essays in biochemistry*, 68(2): 235–251. https://doi.org/10.1042/EBC20230075
- Croft, S. L., Davidson, R. N., & Thornton, E. A. (1991). Liposomal amphotericin B in the treatment of visceral leishmaniasis. *The Journal of antimicrobial chemotherapy*, 28 Suppl B, 111–118. https://doi.org/10.1093/jac/28.suppl\_b.111
- 64. McAuliffe, G. N., Anderson, T. P., Stevens, M., Adams, J., Coleman, R., Mahagamasekera, P., Young, S., Henderson, T., Hofmann, M., Jennings, L. C., & Murdoch, D. R. (2013). Systematic application of multiplex PCR enhances the detection of bacteria, parasites, and viruses in stool samples. *The Journal of infection*, 67(2): 122–129. https://doi.org/10.1016/j.jinf.2013.04.009
- Keddy, K. H., Saha, S., Kariuki, S., Kalule, J. B., Qamar, F. N., Haq, Z., & Okeke, I. N. (2022). Using big data and mobile health to manage diarrhoeal disease in children in low-income and middleincome countries: societal barriers and ethical implications. *The Lancet. Infectious diseases*, 22(5): e130–e142. https://doi.org/10.1016/S1473-3099(21)00585-5
- 66. Ghouneimy, A., Mahas, A., Marsic, T., Aman, R., & Mahfouz, M. (2023). CRISPR-Based Diagnostics: Challenges and Potential Solutions toward Point-of-Care Applications. ACS synthetic biology, 12(1): 1–16. https://doi.org/10.1021/acssynbio.2c00496
- Nixon, S. A., Welz, C., Woods, D. J., Costa-Junior, L., Zamanian, M., & Martin, R. J. (2020). Where are all the anti-helmintics? Challenges and opportunities on the path to new anti-helmintics. International journal for parasitology. *Drugs and drug resistance*, 14: 8–16. https://doi.org/10.1016/j.ijpddr.2020.07.001
- 68. WHO. (2021). Integrated Approaches for NTD and Malaria Control: A Global Perspective. WHO Bulletin, 99: 567-578.
- DeGroote, J. P., Larson, S. R., Zhang, Y., Sugumaran, R. (2012). Application of geospatial technologies for understanding and predicting vector populations and vector-borne disease incidence. *Geography Compass*, 6(11): 645–659. https://doi: 10.1111/gec3.12003
- Poostchi, M., Silamut, K., Maude, R. J., Jaeger, S., & Thoma, G. (2018). Image analysis and machine learning for detecting malaria. *Translational research : the journal of laboratory and clinical*

medicine, 194:

36–55.

https://doi.org/10.1016/j.trsl.2017.12.004

- Siddiqui, A. J., Bhardwaj, J., Saxena, J., Jahan, S., Snoussi, M., Bardakci, F., Badraoui, R., & Adnan, M. (2023). A Critical Review on Human Malaria and Schistosomiasis Vaccines: Current State, Recent Advancements, and Developments. *Vaccines*, *11*(4): 792. https://doi.org/10.3390/vaccines11040792
- 72. Bomford, R. (1989). Adjuvants for anti-parasite vaccines. *Parasitology today (Personal ed.)*, 5(2): 41–46. https://doi.org/10.1016/0169-4758(89)90190-7
- 73. Fenwick A. (2012). The global burden of neglected tropical diseases. *Public health*, *126*(3): 233–236. https://doi.org/10.1016/j.puhe.2011.11.015
- 74. Ssali, A., Pickering, L., Nalwadda, E., Mujumbusi, L., Seeley, J., & Lamberton, P. H. L. (2021). Schistosomiasis messaging in endemic communities: Lessons and implications for interventions from rural Uganda, a rapid ethnographic assessment study. *PLoS neglected tropical diseases*, 15(10): e0009893.

https://doi.org/10.1371/journal.pntd.0009893

- 75. Trienekens, S. C. M., Faust, C. L., Meginnis, K., Pickering, L., Ericsson, O., Nankasi, A., Moses, A., Tukahebwa, E. M., & Lamberton, P. H. L. (2020). Impacts of host gender on *Schistosoma mansoni* risk in rural Uganda-A mixed-methods approach. *PLoS neglected tropical diseases*, 14(5): e0008266. https://doi.org/10.1371/journal.pntd.0008266
- Manzoni, G., Try, R., Guintran, J. O., Christiansen-Jucht, C., Jacoby, E., Sovannaroth, S., Zhang, Z., Banouvong, V., Shortus, M. S., Reyburn, R., Chanthavisouk, C., Linn, N. Y. Y., Thapa, B., Khine, S. K., Sudathip, P., Gopinath, D., Thieu, N. Q., Ngon, M. S., Cong, D. T., Hui, L., ... Tuseo, L. (2024). Progress towards malaria elimination in the Greater Mekong Subregion: perspectives from the World Health Organization. *Malaria journal*, 23(1): 64. https://doi.org/10.1186/s12936-024-04851-z
- 77. Fontes, G., Leite, A. B., de Lima, A. R., Freitas, H., Ehrenberg, J. P., & da Rocha, E. M. (2012). Lymphatic filariasis in Brazil: epidemiological situation and outlook for elimination. *Parasites & vectors*, 5: 272. https://doi.org/10.1186/1756-3305-5-272
- Gething, P. W., Patil, A. P., Smith, D. L., Guerra, C. A., Elyazar, I. R., Johnston, G. L., Tatem, A. J., & Hay, S. I. (2011). A new world malaria map: *Plasmodium falciparum* endemicity in 2010. *Malaria journal*, *10*: 378. https://doi.org/10.1186/1475-2875-10-378
- Ekpunobi, N. F. and Agu, K. C. (2024). Emergence and Re-Emergence of Arboviruses: When Old Enemies Rise Again. Cohesive J Microbiol Infect Dis. 7(2). CJMI. 000658. DOI: 10.31031/CJMI.2024.07.000658.
- Elias, D., Mengistu, G., Akuffo, H., & Britton, S. (2006). Are intestinal helminths risk factors for developing active tuberculosis? *Tropical medicine &*

I

*international health: TM & IH, 11*(4): 551–558. https://doi.org/10.1111/j.1365-3156.2006.01578.x

- Duncan, R., Kourout, M., Grigorenko, E., Fisher, C., & Dong, M. (2016). Advances in multiplex nucleic acid diagnostics for blood-borne pathogens: promises and pitfalls. *Expert review of molecular diagnostics*, *16*(1): 83–95. https://doi.org/10.1586/14737159.2016.1112272
- 82. Ehrenberg, J. P., Zhou, X. N., Fontes, G., Rocha, E. M. M., Tanner, M., & Utzinger, J. (2020). Strategies supporting the prevention and control of neglected tropical diseases during and beyond the COVID-19 pandemic. *Infectious diseases of poverty*, 9(1): 86. https://doi.org/10.1186/s40249-020-00701-7
- 83. Ekpunobi, N., Obidi, N., Okoye, L., Agu, C., Ogunsanya, T., Ogbodo, U., Uwanta, L. (2025). Knowledge and awareness of breast cancer symptoms, risk factors, and early detection methods among Nigerian university students. *World Journal* of Advanced Research and Reviews. 25: 1129-1138. https://doi.org/10.30574/wjarr.2025.25.2.0332
- 84. Neiderud C. J. (2015). How urbanization affects the epidemiology of emerging infectious diseases. *Infection ecology & epidemiology*, 5: 27060. https://doi.org/10.3402/iee.v5.27060