

IJMPR 2023, 7(6), 119-121

International Journal of Modern Pharmaceutical Research

www.ijmpronline.com

SJIF Impact Factor: 5.273

EFFECTS OF NITRATE ON HEMOGLOBIN COUNT, HISTOPATHOLOGY AND MICRONUCLEUS STUDY IN FRESH WATER FISH CATLA CATLA

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Received on: 21/04/2023	ABSTRACT
Revised on: 11/05/2023	Water got contaminated by industrial, agricultural influence and city sewage water.
Accepted on: 31/05/2023	Nitrate present in all industrial and mostly in agricultural waste and also city Sewage
	contain high level of nitrate which directly impact aquatiaquatic life. Nitrate is a major
*Corresponding Author	cause for the formation of met hemoglobin and shortage of hemoglobin which studied
Subhashri Priyadarsini	by cyanmethemoglobin method (1)20 ml of blood samples were taken in Thomas
Research Scholar of Dept. of	pipette at interval of 24, 48 and 72 hours. And nitrate cause stress environment which causes micronucleus and other genetic abnormalities and also a vital histological
Zoology GIET University	change in the gills of fish. 0, 0.5, 1.0 and 2.0 PPM nitrate dodge given to the fish and
Gunupur Odisha.	studied by conducting several tests with two way ANOVA significant variation.
	KEYWORD: Nitrate, Hemoglobin, Histopathology, cyanmethemoglobin method Micronucleus, Genetic modification.

INTRODUCTION

Nitrate is water soluble which highly found in pesticides, fertilizers and industrial waste and mostly in cities swages release high Nitrogen is water soluble substance which highly found in agricultural products level of nitrate into river waters which affect aquatic life. Which adversely change in the aquatic environment and cause a vital genetic modification and physical modification which also cause presence of met hemoglobin Nitrate compounds have been identified as major metabolic products in fish culture. Nitrite may reach toxic concentrations in high density aquaculture systems and in flowing waters due to industrial contamination and fertilizer use. It is an intermediate product in the bacterial oxidation of ammonia to nitrate in conditioned aquaculture systems (Collins, 1975). This nitrogen compound is highly toxic to aquatic organisms and posses a potential threat to cultured fish. Respiratory blood pigment hemoglobin manifests the transport of oxygen. Nitrite an intermediate product of ammonia nitrification, may reach toxic concentration in aquaculture systems when imbalances pour among species of nitrifying Bactria. Nitrite is present at unusually high concentrations in lakes (McCoy, 1972). One physiological response to nitrite is an increase in methemoglobin. The hemoglobin becomes oxidized fe., the ferrous ion (Fe') is oxidized to ferric ion (Fe) and unable to bind and carry molecules of oxygen. Hence, the toxicity of nitrite to fish received much attention in recent years (Russo and Thurston, 1977).

OBJECTIVES

. To find out the effects of Nitrate on hemoglobin count. To find out the effects of Nitrate on gills of Labeo catla fish by Histopathology test.

. To find out genetic modification and Micronucleus formation in stressful environment due to high nitrate content in water.

MATERIAL AND METHODS

. 10-25 cm length with average 500gm weight fishes are used.

. Nitrate with different concentrations which soluble in water.

. 20ml of blood samples were taken in Thomas pipette and cyanmethemoglobin method used for hemoglobin count.

. Histological study done by Microtome method to find out the effects of Nitrate on fish.

. Giemsa stain used for staining in micronucleus Assay.

. Labeo Catla. in the interval of 24, 48, 72hours. Fish blood was smeared on clean and oven dried microscopic slides. These blood smear slides were air dried at 25°C for two hours and then fixed in cold Cortney's fixative for five minutes and were again fixed in methanol for ten minutes and left to air dry at 25°C for 1 h. Slides were stained for M 30 min in 10% aqueous Giemsa and washed in double distilled water and again let them air dry. 35 fish specimens were analyzed for each experimental site for a total of 35,000 erythrocytes/fish sample. For positive control, blood Smith farmed

specimens was subjected to colchicine treatment. For each fish specimen five slides were prepared.

The frequencies of micronucleus induction in erythrocytes were scored.

RESULTS

Hemoglobin concentration (g/dl)

	Α	В	С	D
24hr	5.34±0.02	6.36±0.00	5.36±0.03	6.22±0.01
48hr	5.32±0.01	7.01±0.01	7.14 ± 0.01	5.55±0.13
72hr	5.36±0.02	5.75 ± 0.02	7.18 ± 0.05	5.25 ± 0.01

Fish exposed to different dosage (0.5, 1.0, 2.0ppm)

Hemoglobin concentration of fish in different concentration. Histopathology

	Chloride Cell	Epithelial cell	Lamellar Disruption	Hypertrophy of epithelium
24	00	01	00	01
48	01	00	01	01
72	00	02	01	02

Study showing different abnormal growth of abnormal cancerous cells in gills of fish due to concentration of Nitrate. Micronucleus concentration.

DOSE	24hr	24hr	48hr	48hr	72hr	72hr
	MN	NA	MN	NA	MN	NA
0	0.2	0.3	0.3	0.1	0.2	0.4
0.5ppm	0.3	0.6	0.5	1.1	0.6	0.9
1.0ppm	0.7	1.3	1.3	2.1	1.6	1.7
2.0ppm	1.5	2.1	2.9	3.0	3.3	3.9

MN-Micronucleus NA-Nuclear Anomaly

Frequency-(0/00) Effect of dose and time was studied by conducting the two way ANOVA significant variation between dose(0,0.5,1.0,2.0 ppm) was observed (p<0.01) in micronucleus and nuclear anomaly induction.

RESULTS AND DISCUSSION

The aquatic environment makes up the major part of our environment and resources; therefore, its safety is directly related to the safety of our health and food security. Biomarkers and bio indicators using fish micronucleus assay, histopathology and hemoglobin count process which indicates that with the concentration of Nitrate the damage in fish body increases. Which shows a dangerous condition of aquatic ecosystem due to water pollution.

REFERENCE

- 2007 □Effects of ammonia, nitrite and nitrate on hemoglobin content and oxygenconsumption of freshwater fish, Cyprinus carpio (Linnaeus)K.S. Tilak*, K. Veeraiah and J. Milton Prema Raju Department of Zoology, Acharya Nagarjuna University, Nagarjunanagar – 522 510, India.
- 2. Brown, D.A. and D. McLey: Effect of nitrite on met hemoglobin and total hemoglobin of juvenile rainbrow trout. Pro. Fish Cult., 1975; 3: 36-43.
- 3. Cameron, J.N.: Methemoglobin erythrocytes. Com. Biochem. Physio., 1971; 40: 743-749.

- Collins, M.T.: The effect of nitrite on the short term growth ands urvival of channel catfish, Letarurus purctatus. Aquaculture, 1975; 24: 111-222.. Finney, D.J.: Probit analysis, 3rd Edn. Cambridge University Press (1971). Golteman, H. and Clyma: Methods for the chemical analysis of freshwater. Blackwell Scientific Publications, 1969; 166.
- Hotch kiss, J.H., M.A. Helsen, C.M. Maragos and YM. Weng: Nitrite, nitrate and nitroso compounds in foodsafety assessment (Eds: J.W. Finkey). ACS symposium series, 1992; 484: 400-18.
- Huey, D.W., B.A. Simco and D.W. Cris well: Nitrite induced methe moglobin formation in channel catfish. Trans. Amer. Fish Soc., 1980; 109: 558-562.
- 7. Klinger, K.: Toxicity of nitrite to channel catfis h. Pro. Fish Cult., 1957; 37: 96-98.
- McCoy, E.F.: Role of bacteria in the nitrogen cycle in lakes. Water Pollution Control Res., Ser. /6010 HER. 03/72. Washington DC. Office of Research on Monitoring (1972).
- 9. Perrone, S.J. and T.L. Meade: Protective effect of chloride on nitrite toxicity to Coho solmon. J. Fish Res. Board of Canada, 1977; 34: 486-492.
- Russo, R.C. and R.V. Thurston: The acute toxicity of nitrite to fishes in recent advances in fish toxicology, (Ed: R.A. Tubb). Ecol. Res. SER. EPA 600/3-77-085. Corvallis Ore., USA. Pp.18-131 EPA. (1977).
- 11. Tomasso, J.R.: Comparative toxicity of nitrite to fresh water fishes. Aqua. Toxicol., 1981; 8: 129-137.

- Veeraiah, K. and M.K. Durga Pras ad: Study on the toxic effects of cypermethrin (technic al) on or ganic constituents of fresh water fish Labeo roh it a (Hamilton). Proc. Acad. Environ. Biol., 1998; 7(2): 143-148.
- 13. Toxic effects of chlorpyrifos on fish 5.
- 14. Shi LL, Lin YS, Yu YG, Chen LY. (2000). Studies on environmental behavior of chlorpyrifos pesticide. Soil Environ Sci., 9: 73-74.
- 15. Singh NN, Srivastava AK. (2010). Haematological parameters as bioindicators of insecticide exposure in teleosts. Ecotoxicology, 19: 838-854.
- Starner K, Spurlock F, Gill S, et al. (2005). Pesticide residues in surface water from irrigation-season monitoring in the San Joaquin Valley, California, USA. Bull Environ Contam Toxicol, 74: 920-927.
- 17. Sun F, Chen HS. (2008). Monitoring of pesticide chlorpyrifos residue in farmed fish: investigation of possible sources. Chemosphere, 71: 1866-1869.
- Sun F. Wong S, Li G, Chen S. (2006). A preliminary assessment of consumer's exposure to pesticide residues in fisheries products. Chemosphere, 62: 674-680.
- 19. Svoboda M, Luskova V, Drastichova J, Zlabek V. (2001). The effect of diazinon on haematological indices of common carp (Cyprinus carpio L.). Acta Vet Brno, 70: 457.
- 20. Tavares DM, Martins ML, Nascimento KS. (1999). Evaluation of the haematological parameters in Piaractus mesopotamicus Holmberg (Osteichthyes, Characidae) with Argulus sp. (Crustacea, Branchiura) infestation and treatment with organophosphate. Rev Bras Zool, 16: 553-555.
- U.S. Environmental Protection Agency. (2002). Methods for measuring the acute toxicity of effluents and receiving waters to freshwater and marine organisms. EPA-821-R-02-012. Office of Water Washington eD. CDC.
- 22. Ventura Campos de B, Angelis de DF, Marin-Morales MA. (2008). Mutagenic and genotoxic effects of the Atrazine herbicide in Oreochromis niloticus (Perciformes, Cichlidae) detected by the micronuclei test and the comet assay. Pestic Biochem Physiol, 90: 42-51.
- 23. Yin XH, Zhu GN, Li XB, Liu SY. (2009). Genotoxicity evaluation of chlorpyrifos to amphibian Chinese toad (Amphibian: Anura) by comet assay and micronucleus test. Mutat Res., 680: 2-6.

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