

**PATHOLOGICAL PATHWAY AND PROTECTIVE MECHANISMS OF
PHYTOBIOACTIVES USED IN DIABETIC RETINOPATHY: A REVIEW**Negarish Zia¹, Badruddeen^{1*}, Juber Akhtar¹, Mohammad Irfan Khan¹, Mohammad Ahmad Khan¹, Md. Babar Siddique¹ and Murlyadhar Singh Rathore¹

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Diabetes mellitus (DM) is caused by either inadequate or dysfunctional insulin. It is one leading causes of Diabetic retinopathy (DR). Many synthetic medications have serious adverse effects that exacerbate the diabetic patient's condition. The discovery of phytochemicals from medicinal plants offers a promising possibility for the creation of novel therapies for diabetes and its complications including DR. This review highlighted pathological mechanism of DR and also describe numerous plant-derived small compounds (phytochemicals) that have been studied in pre-clinical and clinical settings for their potential to have retinal cytoprotective effects. This also emphasise the mechanism retinal cryoprotection of phytochemicals. The sources were retrieved from various database such as Science Direct, Google Scholar, PubMed, Medline etc. In various studies, phytochemicals reduced inflammation, apoptosis pathways and oxidative stress, which inhibited the development of DR. Numerous phytochemicals, including flavonoids, lignan, glycosides, phytoestrogen etc., inhibited the production of Reactive Oxygen Species (ROS) and downregulation the inflammatory markers such as Interleukin-6(IL-6), Interleukin-1 β (IL-1 β), and Tumor necrosis factor- α (TNF- α). Overall most of the phytochemicals shows retinal cryoprotection by reducing generation of free radicals and inflammatory mediators.

KEYWORDS: Diabetes, Inflammation, Oxidative stress, Phytochemicals, Retinopathy.**INTRODUCTION**

Retinal illnesses are collectively referred to as retinopathy. They frequently affect the small blood vessels (capillaries) feeding the light-sensitive region of the eye, resulting in injury, fluid leakage, or the creation of new crisp blood vessels.^[1] The sensory membrane that receives light and transforms it into nerve signals is found inside the eye as part of the retina. In the brain, this impulse generates an image. Vision loss could be partial or total. It can appear gradually or unexpectedly, get better on its own or have permanent negative impacts.^[2]

Retinopathy exists in different types, including as:(i) Premature Retinopathy (PR), a fibrous tissue forms behind the lens in this degenerative illness, which is only observed in premature infants and babies with low birth weight. It causes blindness and severe vision impairment.^[3] There are several warning signs, such as crossed eyes, near-sightedness, amblyopia (lazy eye), and retinal detachment.^[4] (ii) Diabetic Retinopathy, high blood glucose levels cause DR by weakening the retina's tiny blood vessels. As a result, the retina produces more fluid, blood, cholesterol, and other lipids, which causes the macula to thicken and expand.^[5] Observing more and

more dark spots, poor night vision, fuzzy vision, seeing colours that seem faded or washed out, losing vision and vision that occasionally shifts from blurry to clear are some consequences.^[6] (iii) Central Serous Retinopathy, triggered by the accumulation of fluid behind the retina, which can seriously impair vision because the retina is made up of a thin tissue layer.^[7] Central vision that is dimmed, blurred, or distorted, straight lines that appear bent, crooked, or irregular in the affected eye, a dark area in the central vision, objects that appear shorter or farther away, and white objects that take on a brownish tinge or seem to duller in colour are all possible symptoms of central serous chorioretinopathy.^[8] (iv) Hypertensive Retinopathy, arises when high blood pressure causes damage to the retinal vessels.^[9] Eye swelling, double vision associated by headaches, blood vessel rupture, and reduced vision, are a few evidence.^[10]

This review explained mainly three pathological pathways of DR which include inflammatory, oxidative stress and apoptosis pathways. This review also emphasises the molecular mechanism of retinal cryoprotection of phytobioactives. These phytochemicals modulate inflammatory, apoptotic and oxidative stress pathways. Numerous phytochemicals, including

flavonoids, lignan, glycosides, phytoestrogen etc., reduces the production of Reactive Oxygen Species and inflammatory markers such as cytokines.

Diabetic Retinopathy

The non-infectious epidemic of the modern world is diabetes mellitus (DM).^[11] The WHO currently recognizes DM as a collection of metabolic illnesses characterized by the existence of hyperglycemia caused by insulin release or dysfunction. Individuals with DM who experience chronic hyperglycemia experience destruction, dysfunctional impairment, and insufficiency to a number of body organs, including the eyes, kidneys, nerves, heart, and blood vessels.^[12] Diabetes mellitus is a multifactorial disease, chronic illness that requires ongoing treatment and seems to have no permanent treatment to date.^[13] Increasing life expectancy in developed countries is one of the reasons why diabetes is more common.^[14] However, a longer longevity in those with diabetes mellitus is also associated with a higher risk of chronic microvascular and macrovascular complications brought on by the disease. Vascular complications are currently the leading cause of death and morbidity in people with diabetes mellitus, despite a dramatic drop in the number of deaths directly attributed to this condition.^[15] One third of diabetic individuals experience diabetic retinopathy, which is the most common, most dreaded by patients, and the leading cause of visual-specific neurovascular consequences of DM over the past 20 years.^[16,17] 80% of blindness in DM patients is triggered by DR,^[18,19] which can strike at any stage of the illness.^[20]

Severe retinal vascular disease known as DR is marked by the development of new blood vessels, increased vascular permeability, and infarction and congestion in the retina.^[21] This is brought on by microangiopathy, which affects the retina's precapillary arteries, capillaries, and venules. Damage arises from internal blood-retinal barrier rupture, which causes microvascular leakage and microvascular occlusion.^[22] DR is eye damage caused by prolonged high blood sugar. The walls of the retinal blood vessels weaken, allowing blood to pass through. This causes swelling and vision problems. Over time, new, weaker blood vessels grow, allowing blood to pass through and covering the center of the retina.^[23] This is because prolonged exposure to high blood sugar damages the retinal capillaries.^[24] Blindness is three times as common in those with diabetes than in the regular populace.^[25] It disrupts the retinal vasculature and results in consequences such as capillary protrusion, congestion, cotton wool patches, aberrant neovascularization, and microaneurysms.^[26]

Sequelae from retinal vascular abnormalities in diabetic retinopathy

Microvascular alterations in the retina can be caused by a chain of disastrous circumstances that are triggered by hyperglycemia. The consistency and functionality of the retina are impacted by these alterations, which eventually

cause vision loss. By the time clinically detectable microvascular modifications can be made, significant and occasionally irreparable harm has already been done. Therefore, monitoring the development of diabetic retinopathy requires early molecular alterations detection.^[27]

Microaneurysms (MA) – These are the first clinically discernible symptoms, structural deterioration and deformation of the capillary wall.^[28] Small sac that can develop as a result of partial capillary wall stretching.^[22] They initially make a brief appearance in the fovea and may later vanish.^[29]

Haemorrhages – Subretinal dot haemorrhages are the consequence of impaired capillary wall punctures. Precapillary arterioles in the retinal nerve fibre layer are the source of superficial or flame-shaped haemorrhages.^[30] The inner nuclear and outer plexiform layers of the retina contain deep haemorrhages, also known as dot and blot haemorrhages, which are more commonly associated with severe hypertension.^[22]

Cotton Wool Spots/Soft Exudate – Cotton wool patches, which are axoplasmic aggregates from nerve fibre terminals, are signs of bloodless tissue in the nerve fibre layer as a result of the closure of retinal micro vessels.^[31] Axonal flow, often referred to as axoplasmic transport proceeds in two directions: among somatic cells (the body of neurons) and synapses which transport organelles and cellular components.^[32]

Retinal oedema – Microvascular leakage causes retinal inflammation, which is a sign that the inner blood-retinal barrier has broken down. It manifests as thickened, greyish regions of the retina. The macula's swelling, which may resemble a petal-shaped cyst, can seriously impair vision.^[22]

IRMA (Intraretinal Microvascular Abnormalities) – It is the linkage between the retinal arterioles and veins that are observable adjacent to the capillary occlusion area and circumvent the capillaries. The retina is where IRMA is found, and it avoids big blood arteries.^[30]

When it refers to diabetic retinopathy, there are two broad categories: Non-Proliferative Diabetic Retinopathy (NPDR) and Proliferative Diabetic Retinopathy (PDR). Despite the fact believed that several, if not all, of the 30 distinct forms of retinal cells are compromised by diabetes, the extent of the vascular lesion determines the type of retinopathy because it is possible to see the inner retinal vasculature.^[33] The main difference between NPDR and PDR is the presence of angiogenesis in PDR.^[20] Macular edema can develop at any time as diabetic retinopathy progresses.^[34]

NPDR can be split into three different phases: minimal, intermediate, and chronic NPDR, depending on the seriousness of the consequences. This stage is a

persistent damage of the retinal microvascular structures that enables diabetic retinopathy grow pathologically.^[35]
 (i) The initial stage is thought to be mild non-proliferative retinopathy, which is linked to the development of MA.
 (ii) Moderate non-proliferative retinopathy, where the retina's blood vessels may expand and deform as the disease progresses, losing their capacity to transport blood.
 (iii) Severe non-proliferative retinopathy causes the retina to receive less blood because more blood vessels are blocked, encouraging the retina to develop new blood vessels. PDR is a highly sophisticated stage of the disease in which the retina's growth factors stimulate the formation of new blood vessels along the retina's inner surface in some vitreous gel that fills the eye.^[36]

Molecular Frameworks Implicated In The Diabetic Retinopathy Pathogenesis

Innumerable pathological parameters, especially hyperglycemia and genetic susceptibility, can trigger the onset and advancement of diabetic retinopathy. In diabetes-induced retinal exertion that eventually result in microvascular destruction and retinopathy, several key systems have been identified, together with (i) the polyol pathway, (ii) soreness, (iii) Redox imbalance, (iv) nonenzymatic glycation, (v) protein kinase C (PKC) excitation, and (vi) congenital. Antecedent in the evolution of proliferative retinopathy, these methodologies boost the expression of VEGF.^[37,38] while also encouraging the emergence of new blood vessels, enhancing vascular permeability, and driving leukocyte activation and adherence (Figure 1).^[39,40]

Persistent hyperglycemia: It is thought to be the predominant pathogenic aspect of DR.^[41] The polyol pathway as well as other additional glucose metabolism

routes are activated by hyperglycemia. Advanced glycation end products (AGEs) are generated as a consequence of PKC activation, non-enzymatic protein glycation and oxidative stress. The stimulation of cytokines, growth factors, and vascular endothelial dysfunction that results from these alternative routes finally causes an increase in vascular permeability and microvascular occlusion. Microvascular blockage triggers retinal ischemia, which encourages the growth of IRMA and neovascularization.^[42]

Redox imbalance: Cell and tissue destruction are the outcome of elevated amounts of reactive oxygen species (ROS).^[43]

The polyol route: The enzyme aldose reductase in this transform's glucose to sorbitol. Because sorbitol is impermeable, it builds up inside all retinal cells, triggering osmotic deterioration of the cells. Additionally, the reduction process's consumption of NADPH (reduced nicotinamide adenine dinucleotide phosphate) caused enormous oxidative destruction.^[44]

PKC: Transduction of signals is involved. Its activation causes abnormalities in the basement membrane and vascular structure, including vascular stasis, capillary occlusion, enhanced vascular permeability, and the generation of angiogenic growth factors.^[45]

Non-enzymatic protein glycation: This causes the production of AGEs, which are in charge of altering the extracellular matrix proteins, when reducing sugars react with lipids, free amino acids of nucleic acids, and proteins.^[46]

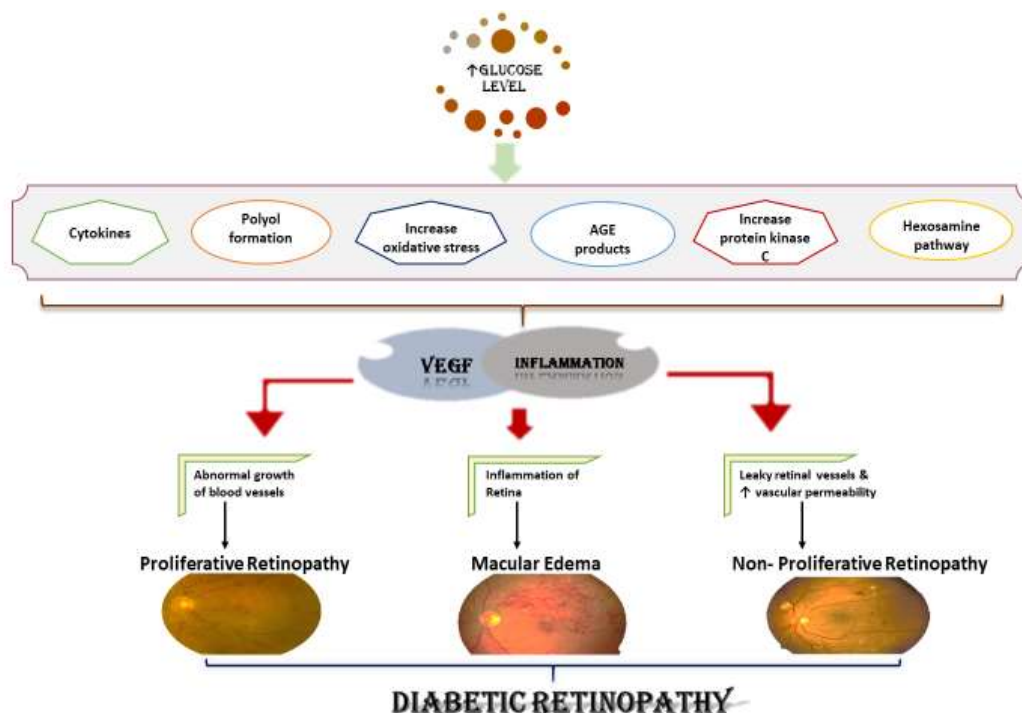


Figure 1: Pathogenesis of DR. (VEGF: Vascular endothelial growth factor).

Phytochemicals For Diabetic Retinopathy Management

Organic natural ingredients called nutraceuticals are enriched with substances including vitamins, antioxidants, minerals, fatty acids, as well as amino acids that can halt the advancement of some disorders or provide a wide range of health improvements. Numerous studies have demonstrated that nutraceuticals provide diverse clinical aspects and defend from a number of ailments.^[47] Nutraceuticals are prescribed to treat diabetes because they improve insulin sensitivity, control metabolism, and lower hyperglycemia.^[48] One of the most widespread and well-known phytonutrients are terpenoids, alkaloids, glycosides, flavonoids, and tannins.^[49] Phytochemicals have well-known antioxidant, antiangiogenic, and anti-inflammatory properties that can be retrieved by food consumption. The majority of phytochemicals are thought to be friendlier treatment options for diabetic retinopathy than pharmaceuticals. In

pre-clinical research, phytochemicals controlled oxidative stress, inflammation, and apoptosis pathways to diminish DR (Figure 2).^[50]

Alkaloids: Alkaloids are secondary plant metabolites that are also present in mammals, microbes, and fungi.^[51] These basic alkaloids typically have heterocyclic structures, contain one or more nitrogen atoms, and are derived from amino acids. Alkaloids are categorised according to their pharmacological properties, chemical make-up, biochemical ancestry, and taxonomic origin. For their possible antidiabetic properties, different alkaloids have been taken from a range of herbal plants and tested on diverse animal species.^[52] It has been suggested that it is one of the active ingredients in some plants used to treat diabetes.^[53] In order to reduce diabetic retinopathy, certain alkaloids include: betaine, cannabidiol, and sinomenine.

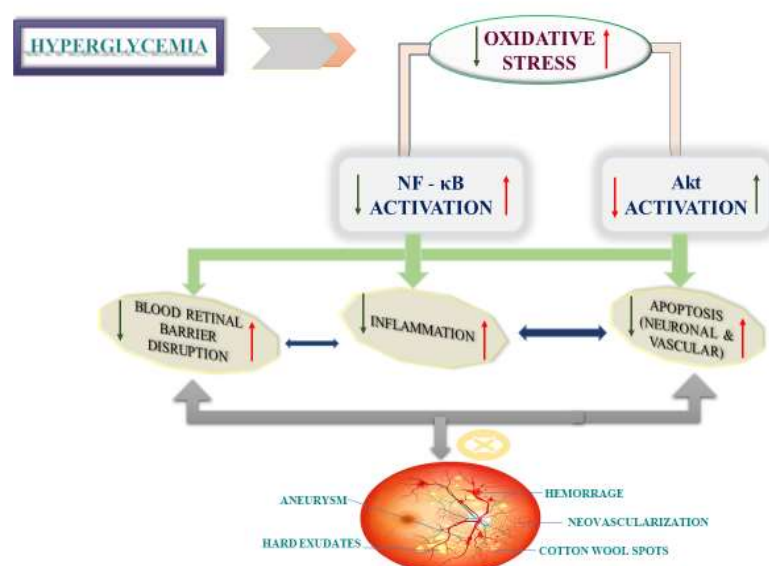


Figure 2: Speculative scenario resulting in diabetic retinopathy from elevated blood glucose levels and the impact of phytochemicals. The green arrow depicts the retardation by phytochemicals, while the red arrow signifies the elevated oxidative stress, inflammation, and apoptotic pathways in DR.

Glycosides: Glycosides are a common type of secondary metabolite found in plants.^[54] The aglycone (genin) and glycone (saccharide) components of glycosides are two chemically and functionally distinct sections. A glycosidic bond connects the saccharide and aglycone portions of a glycoside.^[55] Several glycosides have been shown to diminish diabetic retinopathy, including: arctiin, aloe-emodin, decursin, gastrodin, gentiopicroside, hesperidin, hesperetin, lithospermic acid B, malvidin, paeoniflorin, physcion 8-O-β-glucopyranoside, pterostilbene, sauchinone, scutellarin, and shikonin.

Flavonoids: Among the most abundant and extensively dispersed classes of organic ingredients inside the plant world are polyphenols, which are scientifically referred

to as substances with phenolic structural characteristics. There are many subgroups of phenolic compounds in this group of naturally occurring chemicals, which is very diverse. Fruits and vegetables include flavonoids, a polyphenol subclass with particular biological properties that include anti-inflammatory, antiviral, and antioxidant activities. Flavonoids make up around 60% among all polyphenols.^[56] Flavanols, isoflavones, flavones, anthocyanins, flavanones and flavonols are the six types of flavonoids that can be divided based on their chemical structure.^[57] Flavonoids can regulate lipid and carbohydrate metabolism, alleviate hyperglycemia, better control inflammatory responses, increase insulin resistance, and enhance β-cell performance, which may assist to delay the onset of long-term chronic diabetes consequences including diabetic retinopathy.^[58] The

reduction of diabetic retinopathy by a number of flavonoids has been reported, including: anthocyanins, alpha-mangostin, baicalein, biochanin, curcumin, chrysin, epigallocatechin-3-gallate, eriodictyol, formononetin, genistein, icariin, kaempferol, luteolin, naringin, puerarin, quercetin, resveratrol, rutin, sesamin, silybin, taxifolin, and troxerutin.

Terpenoids: At least 4000 triterpenes are known, and they are produced by the mevalonic acid route. In both plants and animals, terpenes are the building blocks of steroid hormones. These are a group of hydrocarbon substances, such as squalene, that are made up of three terpenes and six isoprene units. Triterpenoids are functionalized triterpenes. One of the largest families of natural products. It also known as "isoprenoids") has more than 40,000 different chemicals that are involved in both primary and secondary metabolism.^[59] These are bioactive substances with strong hypoglycemic effects.^[60] Numerous terpenoids have been shown to dampen the risk of developing diabetic retinopathy, including: andrographolide, astaxanthin, carotenoids,

dammarenediol-II, β , ϵ -Carotene-3,3'-diol, sulforaphane, curcumolide, and zerumbone.

Tannins: Innumerable plant species contains tannins, which are polyphenolic macromolecules with a significant molecular weight. Tannins bind and expel proteins as well as a variety of other chemical compounds like alkaloids and amino acids. Condensed tannins and hydrolysable tannins are the two principal types of tannins. Hydrolysable tannins are composed of polyol (D-glucose), hydroxyl moiety, and phenolic acids such as ellagic acid as well as gallic acid. Condensed tannins are a polyphenolic bioflavonoid called polyhydroxy flavan-3-ol. In current history, tannins have received more attention due to the health benefits associated with their antioxidant capacities.^[61] Certain tannins can minimize diabetic retinopathy, such as: gallic acid, chebulagic acid, chebulinic acid and chlorogenic acid.

The enumeration of alkaloids, glycosides, tannins, flavonoids, and terpenoids along with their origin and mode of action are stated in Table 1.

Table 1: Various phytochemicals effective in DR.

S.no.	Phytochemicals	Source	Mechanism of action	Reference	
1.	Alkaloids	Betaine	Capsicum, Silybum, Beta vulgaris	\downarrow Akt, VEGF, HIF-1 α .	[62]
2.		Cannabidiol	<i>Cannabis sativa</i>	Blockage of p38 MAPK. \downarrow Separation of BBB, ROS, VEGF, TNF- α .	[63]
3.		Sinomenine	<i>Sinomenium acutum</i>	\downarrow microglial development, TNF- α , Inflammation of the retina, IL-1 β , ROS, IL-6, NF- κ B p65.	[64]
4.	Glycosides	Aloe emodin	-	\downarrow HIF-1 α , PHD-2, expressions in retinal neovascularization of VEGFA.	[65]
5.		Arctiin	<i>Arctium lappa L.</i>	\downarrow VEGF, HbA1C. Retinal separation and diminution in retinal edema.	[66]
6.		Decursin	<i>Angelica gigas</i>	\downarrow Ocular neovascularization, VEGFR-2 expression. Retinal proliferation, tube development and angiogenesis of retina.	[67]
7.		Gastrodin	<i>Gastrodia elata</i>	\downarrow SIRT1/TLR4/NF- κ B p65 signaling pathway, NADPH, Nrf2, GCLM, ROS, HO-1 and NQO1, expression of cleaved caspase-3 and cytochrome C. \uparrow Bcl-2/Bax.	[68]
8.		Gentiopicroside	-	\downarrow Oxidative stress, NF- κ B, ICAM-1, IL-1 β , GFAP, TNF- α , MDA, expression	[69]

				of VEGF, protein carbonyl, ROS, HDAC overexpression, inflammation in retina. ↑expression of GSH, CAT, PEDF and SOD.	
9.	Hesperidin	Citrus fruits		↓MDA, AR activity, VEGF, IL-1 β , ICAM-1, TNF- α , oxidative stress, angiogenesis of retina, inflammation. ↑thickness of the retina, expression of SOD and blood retinal barrier permeability.	[70]
10.	Hesperetin	Citrus fruits		↓PKC- β , widened vessels, Vascular permeability, VEGF, stiffening of BM, angiogenesis of retina. ↓ROS, AQP4, caspase-3, inflammation of retina, TNF- α , GFAP, IL-1 β . ↑CAT, SOD and GSH activities in the retina.	[71,72]
11.	Lithospermic acid B	<i>Salvia miltiorrhiza</i>		↑capillary BM coat, nerve layer thickness, and ganglion cells. ↓hsCRP, MCP1, VEGF, 8-OHdG and TNF- α , vascular leakage, and arterial sclerosis of capillary.	[73]
12.	Malvidin	<i>Saccharomyces cerevisiae</i>		↓Nox4 activity, Mv-3-glc, Mv-3-gal, and Mv.	[74]
13.	Paeoniflorin	<i>Paeonia lactiflora</i>		↓MMP-9 activation, inflammation of retina, translocation of NF- κ B p65, p-p38 expression, IL-1 β , IBA-1. ↑activation of TLR4, thickness of retina, expression of SOCS3 and gliocyte proliferation of retina.	[75]
14.	Physcion8-O- β -glucopyranoside	<i>Rheum palmatum</i> , <i>Rheum australe</i> , and <i>Senna obtusifolia</i>		↓expression of STAT3 and NORAD, generation of ROS, IL-1 β , TNF- α , apoptosis of cell. ↑expression of miR-125.	[76]
15.	Pterostilbene	<i>Vitis rupestris</i> , <i>Pterocarpus marsupium</i>		↓Production of ROS, TNF- α , IL-1 β , mRNA, expression of protein and NF- κ B. ↑activation of SOD.	[77]
16.	Sauchinone	<i>Saururus chinensis</i>		↓Bcl-2, ROS. ↑Akt/Nrf2/HO-1 signaling pathway, CAT, Bax, GPx, SOD.	[78]
17.	Scutellarin	<i>Scoparia dulcis</i> , <i>Sempervivum</i>		↓VEGF, activity of NADPH oxidase, ROS,	[79,80]

			<i>ruthenicum</i>	HIF-1 α , ERK, FAK, p-Src phosphorylation, proliferation and angiogenesis of retina and development of tube.	
18.		Shikonin	<i>Lithospermum erythrorhizon</i>	↓ZO-1, iNOS, MPO, Bax, COX-2, inflammation of retina, damage of retinal cell, vascular permeability, edema.	[81]
19.	Flavonoids	Anthocyanins	<i>Vaccinium myrtillus</i> , <i>Vaccinium virgatum</i>	↓ROS, VEGF, tight junction proteins loss, breakdown of blood retina barrier, Mv, MV-3-gal, Mv-3-glc, BAE, Akt. ↑SOD, CAT.	[82,74]
20.		α -mangostin	<i>Garcinia mangostana</i>	↓MDA, TNF- α , VEGF	[83]
21.		Baicalein	<i>Scutellaria baicalensis</i>	↓VEGF, IL-1 β , TNF- α , IL-18, GFAP, inflammation of retina, GLC loss, activation of microglial cell, vascular permeability	[84]
22.		Biochanin	<i>Trifolium pratense</i>	↓VEGF, ICAM-1, TNF- α , IL-1 β , inflammation, angiogenesis of retina. ↑NF- κ B.	[85]
23.		Curcumin	<i>Curcuma longa</i>	↓VEGF, width of the retinal capillary BM, retinal angiogenesis, 8-OHdG, nitro tyrosine, TNF- α , diameter of vessel, MDA, NF- κ B phosphorylation, CaMKII, iNOS, and ICAM-1 expressions, inflammation of retina, IL-1 β , HbA1c, vascular leakage of retina, oxidative stress, IL-6, ROS-AKT/mTOR, GFAP. ↑GSH, SOD, T-AOC, CAT, Brn3, Ratio of Bcl-2 to Bax, RecA, Thy-1.	[86-91]
24.		Chrysin	<i>Passiflora caerulea</i>	↓VEGF, IGF-1, secretion of AGE, RAGE, HIF-1 α , Ang- Tie-2 pathway, neovascularization of retina, ER stress. ↑Thickness of retina, RPE65, PECAM-1, PEDF, RDH5, LRAT, ZO-1 and VE- cadherin junction proteins.	[92,93]
25.		Epigallocatechin-3-Gallate	<i>Camellia sinensis</i> L.	↓ERK1/2, MAPK, VEGF.	[94]

26.	Eriodictyol	<i>Eriodictyon californicum</i>	↓oxidative stress, eNOS, VEGF, TNF- α , CAM-1, ROS, IL8, inflammation of retina, BAX, cleaved caspase-3. ↑CAT, SOD, GPX, Bcl-2, Nrf2/HO-1 activation.	[95]
27.	Formononetin	<i>Astragalus membranaceus-s</i>	↓VEGF, HIF- α , neovascularization of retina.	[96]
28.	Genistein	<i>Glycine max</i>	↓VEGF165 and its secretion, ALR, ROS, TNF- α , retinal microglial cell activation, oxidative stress, P38 MAPKs and ERK activation, retinal inflammation and angiogenesis.	[97]
29.	Icariin	<i>Epimedii Herba</i>	↓VEGF, RECA. ↑Brn3a, Thy-1.	[98]
30.	Kaempferol	-	↓VEGF, PGF, retinal angiogenesis, Erk1/2, Src, Akt1, P13K expression.	[99]
31.	Luteolin	<i>Platycodon grandiflorus</i>	↓VEGF, IL-1 β , NF- κ B.	[100]
32.	Naringin	-	↓GFAP level, IL-1 β , IL-6, TNF- α , NF- κ B p65, oxidative stress, retinal inflammation. ↑GSH, SOD, thickness and cell number of ganglions.	[101]
33.	Puerarin	<i>Pueraria montana, Radix Puerariae</i>	↓ROS, NADPH oxidase activity, Rac1, p47phox, NF- κ B, 8-OHdG, VEGF, HIF- α .	[102,103]
34.	Quercetin	-	↓MCP-1, ROS, IL-6, apoptosis of cell, NF- κ B. ↑miR-29b overexpression.	[104]
35.	Resveratrol	Grapes, and berries	↓oxidative stress, NF- κ B, IL-6, TNF- α , COX-2, apoptosis, cell death, basement membrane thickness, vascular hyperpermeability, eNOS, ACE, MM-9 expression.	[105]
36.	Rutin	Onions, Apples, Tea and Red wine	↓Pro- apoptotic pathway, neuronal apoptosis. ↑NGF, BDNF.	[106]
37.	Sesamin	<i>Sesamum indicum</i>	↓TNF- α , microglial activation, ICAM-1, iNOS, ROS.	[107]
38.	Silybin	<i>Silybum marianum</i>	↓ICAM-1, Retinal vascular leukostasis, retinal capillary deterioration.	[108]
39.	Taxifolin	-	↓tGSH level, MDA, oxidative stress, TNF- α , IL-1 β .	[109]

40.		Troloxerutin	<i>Sophora japonica</i>	↓VEGF, oxidative stress.	[110]
41.	Terpenoids	Andrographolide	<i>Andrographis paniculata</i>	↓VEGF, TF, retinal angiogenesis, I-κK, I-κB, NF-κBp65, retinal inflammation, Egr-1, TNF-α, IL-6, IL-1β.	[111]
42.		Astaxanthin	Carotenoids present in plants, algae and seafood	↓Oxidative stress, anti-apoptosis pathways.	[112]
43.		Carotenoids	<i>β-carotene</i>	↓Oxidative stress, VEGF, ICAM-1, LPO.	[113]
44.		Curcumolide	<i>Curcuma wenyujin</i>	↓ICAM-1, retinal vascular leakage, leukostasis, p38 MAPK, TNF-α, NF-κB.	[114]
45.		Dammarenediol-II	<i>Panax ginseng</i>	↓VEGF, ROS, formation of stress fibre, microvascular leakage in retina, breakage of vascular endothelial-cadherin.	[115]
46.		β, ε-Carotene-3,3'-diol	-	↓Nitro tyrosine level. ↑MDA, GSH, GPx.	[116]
47.		Sulforaphane	-	↓Oxidative stress, inflammation, formation of tumour, TNF-α, IL-1β, IL-6, NLRP3, cleaved caspase-1p20, ASC level. ↑GSH, CAT, SOD, ganglion cells count, NQO1, HO-1, Nrf2.	[117]
48.		Zerumbone	<i>Zingiber zerumbet</i>	↓Nerve fibres layer, retinal thickness, ganglion cells, IL-6, IL-1β, TNF-α, RAGE, VEGF, NF-κB, VCAM-1.	[118]
49.	Tannins	CA, CI, and GA	-	↓Retinal angiogenesis, MMP-9 expression, TNF-α, p38, NF-κB, ERK, IL-6, MCP-1, IL-8, RANTES, MIP-1b, eotaxin. ↑IL-13, IL-10.	[119]
50.		Chlorogenic acid	-	↓VEGFR2, ERK1/2, VEGF, MEK1/2, activity of microglia cell, p38, retinal neovascularization.	[120]

Abbreviations: ↓: Decrease, ↑: Increase, Akt: protein kinase B-1, VEGF: Vascular endothelial growth factor, HIF-1α: Hypoxia-inducible factor-1α, MAPK: Mitogen-activated protein kinases, BBB: Blood retinal barrier, ROS: Reactive oxygen species, TNF-α: Tumor necrosis factor-α, IL: Interleukin, NFκB: Nuclear factor kappa-light-chain-enhancer of activated B cells, PHD-2: prolyl hydroxylase domain protein 2, HbA1 C: Glycosylated hemoglobin, VEGFR-2: vascular endothelial growth factor receptor, SIRT1: sirtuin 1, TLR4: Toll-like

receptor 4, NADPH: nicotinamide adenine dinucleotide phosphate, Nrf2: nuclear factor erythroid 2-related factor 2, GCLM: c-glutamate-cysteine ligase modifier, HO-1: heme-oxygenase-1, NQO1: NADPH quinone oxidoreductase 1, Bcl-2: B-cell lymphoma 2, Bax: BCL-associated X, ICAM-1: Intercellular adhesion molecule-1, GFAP: Glial fibrillary acidic protein, MDA: Malondialdehyde, HDAC: Histone deacetylases, GSH: glutathione peroxidase, SOD: superoxide dismutase, PEDF: pigment epithelium derived factor, AR: aldose

reductase, PKC β : protein kinase C β , BM: basement membrane, AQP4: aquaporin-4, GFAP: glial fibrillary acidic protein, CAT: catalase, hsCRP: high sensitivity C-reactive protein, MCP-1: Monocyte chemoattractant protein-1, 8-OHdG: 8-hydroxy-2-deoxyguanosine, Nox4: NADPH oxidase 4, Mv-3-glc: malvidin-3-glucoside, Mv-3-gal: malvidin-3-galactoside, Mv: Malvidin, MMP-9: matrix metalloproteinase-9, p-p38: Phosphorylated-p38 mitogen-activated protein kinase, Iba-1: Ionized calcium-binding adapter molecule 1, SOCS3: suppressor of cytokine signaling 3, STAT3: signal transducer and activator of transcription 3, NORAD: Non-Coding RNA Activated By DNA Damage, miR-125: MicroRNA-125, GPx: glutathione peroxidase, ERK: Extracellular signal-regulated kinase, FAK: focal adhesion kinase, p-Src: Proto-oncogene tyrosine-protein kinase Src, ZO-1: zonula occludens 2, iNOS: Inducible nitric oxide synthase, MPO: myeloperoxidase, COX-2: cyclooxygenase-2, BAE: blueberry anthocyanin extract, GLC: ganglion cell layer, CaMKII : Calcium/calmodulin-dependent protein kinase II, mTOR : mammalian target of rapamycin, T-AOC: total antioxidant capacity, IGF-1: insulin-like growth factor-I, AGE: advanced glycation end, RAGE: Receptor for Advanced Glycation End products, ER: endoplasmic reticulum, RPE: retinal pigment epithelium, PECAM-1: platelet endothelial cell adhesion molecule-1, RDH5: retinol dehydrogenase 5, LRAT: lecithin retinol acyl transferase, VE-cadherin: Vascular endothelial cadherin, eNOS: Endothelial NOS, ALR: aldose reductase, PGF: placenta growth factor, PI3K : phosphoinositide 3-kinases, NGF: nerve growth factor, BDNF: brain-derived neurotrophic factor, I κ B: inhibitor of kappa B, I κ K: inhibitor of kinase, Egr1: Early growth response-1, LPO: lipid peroxidation, MAPK: mitogen-activated protein kinase, NLRP3: pyrin domain-containing 3, ASC: adaptor protein apoptosis associated speck-like protein, VCAM-1: vascular cell adhesion molecule-1, MCP-1: monocyte chemoattractant protein-1, MIP-1b: macrophage inflammatory protein-1, RANTES: regulated upon activation, normal T cell expressed and secreted, MEK: mitogen-activated extracellular regulated kinase.

CONCLUSION

The published research showed the evidences of the application of medicinal plants, either as the source of specific extracted constituents or as a combination of various bioactive substances, which exhibits impressive mitigation of cellular damage to the retina or the enhancement of vision.

As per research studies, phytochemicals reduced inflammation, apoptosis pathways and oxidative stress, which inhibited the development of DR. Numerous phytochemicals, including flavonoids, lignan, polyphenols, iridoid glycosides, pyranocoumarin, xanthoid, anthraquinone, sesquiterpene, naphthoquinone, anthocyanins, isothiocyanate, monoterpene glycoside, and isoquinoline, phytoestrogen, inhibited the production of ROS, angiogenic factors, such as PKC β , HIF-1 α and

VEGF, and the activity of antioxidant enzyme including MDA, SOD, CAT and NADPH oxidase. Additionally, phytochemicals were found to downregulate the inflammatory markers IL-6, IL-1 β , and TNF- α , which have been shown to harm the retina.

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CONFLICT OF INTEREST

The authors declared for none conflict of interest.

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REFERENCES

1. Maxine Lipner, Types of Retinopathy. Verywellhealth Website.
2. <https://www.verywellhealth.com/types-of-retinopathy-5208745>. Published December 06, 2021. Accessed October 11, 2022.
3. Retinopathy Guide: Causes, Symptoms & Treatment options. Drugs.com Website. <https://www.drugs.com/health-guide/retinopathy.html>. Accessed October 11, 2022.
4. Gilbert C, Rahi J, Eckstein M, O'Sullivan J, Foster A. Retinopathy of prematurity in middle-income countries. *Lancet*, 1997; 350: 12-14.
5. Retinopathy of Prematurity. National Eye Institute Website.
6. <https://www.nei.nih.gov/learn-about-eye-health/eye-conditions-and-diseases/retinopathy-prematurity>. Updated June 24, 2022. Accessed January 2, 2023.
7. H. H. Vo and A. Verma. Discriminant color texture descriptors for diabetic retinopathy recognition. *IEEE 12th International Conference on Intelligent Computer Communication and Processing (ICCP)*, 2016; 309-315.
8. Kierstan Boyd. Diabetic Retinopathy: Causes, Symptoms, Treatment. American Academy of Ophthalmology Website.
9. <https://www.aao.org/eye-health/diseases/what-is-diabetic-retinopathy>. Updated October 27, 2022. Accessed January 4, 2023.
10. Jenna Fletcher. Central Serous Retinopathy. MedicalNewsToday Website. <https://www.medicalnewstoday.com/articles/320606>. Updated January 13, 2018. Accessed December 25, 2022.
11. Daniel Porter. What is Central Serous Chorioretinopathy. American Academy of Ophthalmology Website.
12. <https://www.aao.org/eye-health/diseases/what-is-central-serous-retinopathy>. Updated Sep. 21, 2022. Accessed January 4, 2023.

13. Kabedi NN, Mwanza JC, Lepira FB, Kayembe TK, Kayembe DL. Hypertensive retinopathy and its association with cardiovascular, renal and cerebrovascular morbidity in Congolese patients. *Cardiovascular Journal of Africa*, 2014; 25(5): 228-32.
14. Chitra Badii. Hypertensive Retinopathy: Symptoms, Causes, Risk factors, Diagnosis, Types, Complications, Treatments, Outlook, Prevention. Healthline Website.
15. <https://www.healthline.com/health/hypertensive-retinopathy>. Updated February 12, 2021. Accessed, January 7, 2023.
16. Matuszewski W, Baranowska-Jurkun A, Stefanowicz-Rutkowska MM, Gontarz-Nowak K, Gątorska E, Bandurska-Stankiewicz E. The safety of pharmacological and surgical treatment of diabetes in patients with diabetic retinopathy—a review. *Journal of Clinical Medicine*, 2021; 10(4): 705.
17. World Health Organization. Definition, diagnosis and classification of diabetes mellitus and its complications: report of a WHO consultation. Part 1, Diagnosis and classification of diabetes mellitus. *World Health Organization*, 1999; 59.
18. Silva PS, Cavallerano JD, Sun JK, Aiello LM, Aiello LP. Effect of systemic medications on onset and progression of diabetic retinopathy. *Nature Reviews Endocrinology*, 2010; 6(9): 494-508.
19. Kollias AN, Ulbig MW. Diabetic retinopathy: early diagnosis and effective treatment. *Deutsches Arzteblatt International*, 2010; 107(5): 75-84.
20. Centers for Disease Control and Prevention. National diabetes fact sheet: general information and national estimates on diabetes in the United States, 2007.
21. Kempen JH, O'Colmain BJ, Leske MC, Haffner SM, Klein R, Moss SE, Taylor HR, Hamman RF. The prevalence of diabetic retinopathy among adults in the United States. *Archives of ophthalmology (Chicago, Ill.: 1960)*, 2004; 122(4): 552-63.
22. Solomon SD, Chew E, Duh EJ, Sobrin L, Sun JK, Vander Beek BL, Wykoff CC, Gardner TW. Diabetic retinopathy: A position statement by the American Diabetes Association. *Diabetes care*, 2018; 40(3): 412-8.
23. Aiello, L.P.; Gardner, T.W.; King, G.L. Diabetic retinopathy. *Diabetes Care*, 1998; 21: 143-156.
24. Solomon, S.D., Chew, E., Duh, E.J., Sobrin, L., Sun, J.K., Van der Beek, B.L., Wykoff, C.C., Gardner, T.W. Diabetic Retinopathy: A Position Statement by the American Diabetes Association. *Diabetes Care*, 2017; 40: 412-418.
25. Le HG and Shakoor A. Diabetic and Retinal Vascular Eye Disease. *Medical Clinics*, 2021; 105(3): 455-472.
26. Do DV, Wang X, Vedula SS, Marrone M, Sleilati G, Hawkins BS, Frank RN. Blood pressure control for diabetic retinopathy. *Cochrane Database Systematic Review*, 2015; 1: CD006127.
27. Watkins PJ. Retinopathy. *BMJ.*, 2003; 326(7395): 924-6.
28. Diabetic Retinopathy. Drugs.com Website. <https://www.drugs.com/cg/diabetic-retinopathy.html>. Updated February 06, 2023. Accessed February 10, 2023.
29. Antonetti DA, Klein R, Gardner TW. Diabetic retinopathy. *New England Journal of Medicine*, 2012; 366(13): 1227-39.
30. Hayward, L.M., Burden, M.L., Burden, A.C., Blackledge, H., Raymond, N.T., Botha, J.L., Karwatowski, W.S.S., Duke, T. and Chang, Y.F. What is the prevalence of visual impairment in the general and diabetic populations: are there ethnic and gender differences? *Diabetic medicine*, 2002; 19(1): 27-34.
31. Beltramo E, Porta M. Pericyte loss in diabetic retinopathy: Mechanisms and consequences. *Current Medicinal Chemistry*, 2013; 20(26): 3218-3225.
32. Wright WS, Eshaq RS, Lee M, Kaur G, Harris NR. Retinal Physiology and Circulation: Effect of Diabetes. *Comprehensive Physiology*, 2020; 10(3): 933-974.
33. Frank RN. *Etiologic mechanisms in diabetic retinopathy*, Chapter 66. In: Ryan SJ, Hinton DR, Schachat AP, Wilkinson CP, editors. *Retina* (4th ed). Philadelphia, PA: Elsevier Mosby, 2006; 1241-1270.
34. Ezra E, Keinan E, Mandel Y, Boulton ME, Nahmias Y. Non- dimensional analysis of retinal microaneurysms: critical threshold for treatment. *Integrative Biology*, 2013; 5(3): 474-80.
35. Shukla UV, Tripathy K. Diabetic Retinopathy. *StatPearls [Internet]*, 2022. Treasure Island (FL): StatPearls.
36. McLeod D. Why cotton wool spots should not be regarded as retinal nerve fibre layer infarcts. *British Journal of Ophthalmology*, 2005; 89(2): 229-37.
37. McLeod D, Marshall J, Kohner EM, Bird AC. The role of axoplasmic transport in the pathogenesis of retinal cotton-wool spots. *British Journal of Ophthalmology*, 1977; 61(3): 177-191.
38. Stitt AW, Curtis TM, Chen M, Medina RJ, McKay GJ, Jenkins A, Gardiner TA, Lyons TJ, Hammes HP, Simó R, Lois N. The progress in understanding and treatment of diabetic retinopathy. *Progress in retinal and eye research*, 2016; 51: 156-86.
39. Optometric clinical practice guideline: care of the patient with diabetes mellitus. *Eye care of the patient with diabetes mellitus (2nd edition)*. American Optometric Association, 2002.
40. Fong, D.S., Aiello, L., Gardner, T.W., King, G.L., Blankenship, G., Cavallerano, J.D., Ferris III, F.L., Klein, R. Retinopathy in diabetes. *Diabetes care*, 2006; 27(suppl_1): s84-s87.

41. R. Williams. Epidemiology of diabetic retinopathy and macular oedema: a systematic review, *Eye*, 2004; 18(10): 963-983.
42. Adamis AP, Miller JW, Bernal MT, D'Amico DJ, Folkman J, Yeo TK, Yeo KT. Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. *American Journal of Ophthalmology*, 1994; 118(4): 445-450.
43. Boulton M, Foreman D, Williams G, McLeod D. VEGF localisation in diabetic retinopathy. *British Journal of Ophthalmology*, 1998; 82(5): 561-568.
44. Amin RH, Frank RN, Kennedy A, Elliott D, Puklin JE, Abrams GW. Vascular endothelial growth factor is present in glial cells of the retina and optic nerve of human subjects with nonproliferative diabetic retinopathy. *Investigate Ophthalmology & Visual Science*, 1997; 38(1): 36-47.
45. Ishida S, Usui T, Yamashiro K, Kaji Y, Ahmed E, Carrasquillo KG, Amano S, Hida T, Oguchi Y, Adamis AP. VEGF164 is proinflammatory in the diabetic retina. *Investigate Ophthalmology & Visual Science*, 2003; 44(5): 2155-2162.
46. Nathan DM, Genuth S, Lachin J, Cleary P, Crofford O, Davis M, Rand L, Siebert C. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *The New England journal of medicine*, 1993; 329(14): 977-86.
47. Behl T, Kotwani A. Exploring the various aspects of the pathological role of vascular endothelial growth factor (VEGF) in diabetic retinopathy. *Pharmacology Research*, 2015; 99: 137-48.
48. Kern TS, Antonetti DA, Smith LEH. Pathophysiology of Diabetic Retinopathy: Contribution and Limitations of Laboratory Research. *Ophthalmic Research*, 2019; 62(4): 196-202.
49. Wang W, Lo ACY. Diabetic Retinopathy: Pathophysiology and Treatments. *International Journal of Molecular Science*, 2018; 19(6): 1816.
50. Koya D, King GL. Protein kinase C activation and the development of diabetic complications. *Diabetes*, 1998; 47(6): 859-866.
51. Brownlee M. Advanced protein glycosylation in diabetes and aging. *Annu Rev Med*, 1995; 46: 223-234.
52. Sharma, R., Amin, H., Prajapati, P.K. Plant kingdom Nutraceuticals for diabetes. *Journal of Ayurvedic & Herbal Medicine*, 2016; 2(6): 224-228.
53. Matos AL, Bruno DF, Ambrósio AF, Santos PF. The Benefits of Flavonoids in Diabetic Retinopathy. *Nutrients*, 2020; 12(10): 3169.
54. Pathania S, Ramakrishnan SM, Bagler G. Phytochemica: a platform to explore phytochemicals of medicinal plants. *Database (Oxford)*, 2015.
55. Surveswaran S, Zhong Cai Y, Corke H, Sun M. Systematic evaluation of natural phenolic antioxidants from 133 Indian medicinal plants. *Food Chemistry*, 2007; 102(3): 938-953.
56. William J. *Invitation to Organic Chemistry*. Jones and Bartlett, 1999; 433.
57. Belyagoubi-Benhammou, N.; Belyagoubi, L.; Gismondi, A.; di Marco, G.; Canini, A.; Atik Bekkara, F. GC/MS Analysis, and Antioxidant and Antimicrobial Activities of Alkaloids Extracted by Polar and Apolar Solvents from the Stems of *Anabasis Articulata*. *Medicinal Chemistry Research*, 2019; 28: 754-767.
58. Aba, Patrick Emeka, and Isaac Uzoma Asuzu. Mechanisms of actions of some bioactive anti diabetic principles from phytochemicals of medicinal plants: A review. *NISCAIR-CSIR*, 2018.
59. Ikan R. *Naturally Occurring Glycosides*. Chichester, England: John Wiley & Sons Ltd, 1999.
60. Bartnik M, Facey PC. Glycosides. *In Pharmacognosy*. Academic Press, 2017; 101-161.
61. Neveu, V., Perez-Jiménez, J., Vos, F., Crespy, V., du Chaffaut, L., Mennen, L., Knox, C., Eisner, R., Cruz, J., Wishart, D. and Scalbert, A. Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. *Database*, 2010.
62. David, A.V.A., Arulmoli, R. and Parasuraman, S. Overviews of biological importance of quercetin: A bioactive flavonoid. *Pharmacognosy reviews*, 2016; 10(20): 84.
63. Lin, D., Xiao, M., Zhao, J., Li, Z., Xing, B., Li, X., Kong, M., Li, L., Zhang, Q., Liu, Y. and Chen, H. An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. *Molecules*, 2016; 21(10): 1374.
64. Goto T, Takahashi N, Hirai S, Kawada T. Various Terpenoids Derived from Herbal and Dietary Plants Function as PPAR Modulators and Regulate Carbohydrate and Lipid Metabolism. *PPAR Research*, 2010; 483958.
65. Rao, A.V. & Gurfinkel, D.M. The bioactivity of saponins: triterpenoid and steroidal glycosides. *Drug Metabolism & Drug Interaction*, 2000; 17(1-4): 211-235.
66. Laddha AP & Kulkarni YA. Tannins and vascular complications of Diabetes: An update. *Phytomedicine*, 2019; 56: 229-245.
67. Kim YG, Lim H.H, Lee S.H, Shin M.S, Kim C.J, Yang HJ. Betaine inhibits vascularization via suppression of Akt in the retinas of streptozotocin-induced hyperglycemic rats. *Molecular Medicine Reports*, 2015; 12(2): 1639-1644.
68. El-Remessy AB, Al-Shabrawey M, Khalifa Y, Tsai NT, Caldwell RB, Liou GI. Neuroprotective and blood-retinal barrier-preserving effects of cannabidiol in experimental diabetes. *The American journal of pathology*, 2006; 168(1): 235-244.

69. A. L. Wang, Z. Li, M. Yuan, A. C. H. Yu, X. A. Zhu, and M. O. M. Tso. Sinomenine inhibits activation of rat retinal microglia induced by advanced glycation end products. *International Immunopharmacology*, 2007; 7(12): 1552-1558.
70. Wu, J., Ke, X., Wang, W., Zhang, H., Ma, N., Fu, W., Zhao, M., Gao, X., Hao, X. and Zhang, Z., 2016. Aloe-emodin suppresses hypoxia-induced retinal angiogenesis via inhibition of HIF-1 α /VEGF pathway. *International Journal of Biological Sciences*, 2016; 12(11): 1363.
71. Lu LC, Zhou W, Li ZH, Yu CP, Li CW, Luo MH, Xie H. Effects of arctiin on streptozotocin-induced diabetic retinopathy in Sprague-Dawley rats. *Planta Medica*, 2012; 78(12): 1317-1323.
72. Yang, Y., Yang, K., Li, Y., Li, X., Sun, Q., Meng, H., Zeng, Y., Hu, Y. and Zhang, Y., 2013. Decursin inhibited proliferation and angiogenesis of endothelial cells to suppress diabetic retinopathy via VEGFR2. *Molecular and cellular endocrinology*, 2013; 378(1-2): 46-52.
73. T. H. Zhang, C. M. Huang, X. Gao, J. W. Wang, L. L. Hao, and Q. Ji. Gastrodin inhibits high glucose-induced human retinal endothelial cell apoptosis by regulating the SIRT1/TLR4/NF- κ B signaling pathway. *Molecular Medicine Reports*, 2018; 17(6): 7774-7780.
74. X. Zhang, E. Shi, L. Yang, W. N. Fu, F. Hu, and X. S. Zhou. Gentiopicroside attenuates diabetic retinopathy by inhibiting inflammation, oxidative stress, and NF- κ B activation in rat model. *European Journal of Inflammation*, 2019; 17.
75. Shi, X., Liao, S., Mi, H., Guo, C., Qi, D., Li, F., Zhang, C. and Yang, Z., 2012. Hesperidin prevents retinal and plasma abnormalities in streptozotocin-induced diabetic rats. *Molecules*, 2012; 17(11): 12868-12881.
76. B. Kumar, S. K. Gupta, B. P. Srinivasan, T. C. Nag, S. Srivastava, and R. Saxena. Hesperetin ameliorates hyperglycemia induced retinal vasculopathy via anti-angiogenic effects in experimental diabetic rats. *Vascular Pharmacology*, 2012; 57(5-6): 201-207.
77. Kumar, B., Gupta, S.K., Srinivasan, B.P., Nag, T.C., Srivastava, S., Saxena, R. and Jha, K.A. Hesperetin rescues retinal oxidative stress, neuroinflammation and apoptosis in diabetic rats. *Microvascular Research*, 2013; 87: 65-74.
78. Jin, C.J., Yu, S.H., Wang, X.M., Woo, S.J., Park, H.J., Lee, H.C., Choi, S.H., Kim, K.M., Kim, J.H., Park, K.S. and Jang, H.C., 2014. The effect of lithospermic acid, an antioxidant, on development of diabetic retinopathy in spontaneously obese diabetic rats. *PLoS One*, 2014; 9(6): e98232.
79. W. Huang, Z. Yan, D. Li, Y. Ma, J. Zhou, and Z. Sui. Antioxidant and anti-inflammatory effects of blueberry anthocyanins on high glucose-induced human retinal capillary endothelial cells. *Oxidative Medicine and Cellular Longevity*, 2018.
80. Zhu, S.H., Liu, B.Q., Hao, M.J., Fan, Y.X., Qian, C., Teng, P., Zhou, X.W., Hu, L., Liu, W.T., Yuan, Z.L. and Li, Q.P. Paeoniflorin suppressed high glucose-induced retinal microglia MMP-9 expression and inflammatory response via inhibition of TLR4/NF- κ B pathway through upregulation of SOCS3 in diabetic retinopathy. *Inflammation*, 2017; 40: 1475-1486.
81. Wan, W., Wan, W., Long, Y., Li, Q., Jin, X., Wan, G., Zhang, F. and Lv, Y. RETRACTED ARTICLE: physcion 8-O- β -glucopyranoside exerts protective roles in high glucose-induced diabetic retinopathy via regulating lncRNA NORAD/miR-125/STAT3 signalling. *Artificial Cells, Nanomedicine, and Biotechnology*, 2020; 48(1): 463-472.
82. H. Shen and H. Rong. Pterostilbene impact on retinal endothelial cells under high glucose environment. *International Journal of Clinical and Experimental Pathology*, 2015; 8(10): 12589-12594.
83. Y. Shi, Y. Zhang, Y. Li, and C. Tong. Sauchinone inhibits high glucose-induced oxidative stress and apoptosis in retinal pigment epithelial cells. *RSC Advances*, 2021; 9(30): 17065-17071.
84. Long, L., Li, Y., Yu, S., Li, X., Hu, Y., Long, T., Wang, L., Li, W., Ye, X., Ke, Z. and Xiao, H. Scutellarin prevents angiogenesis in diabetic retinopathy by downregulating VEGF/ERK/FAK/Src pathway signaling. *Journal of diabetes research*, 2019.
85. Wang, D., Wang, L., Gu, J., Yang, H., Liu, N., Lin, Y., Li, X. and Shao, C. Scutellarin Inhibits High Glucose-induced and Hypoxia-mimetic Agent-induced Angiogenic Effects in Human Retinal Endothelial Cells through Reactive Oxygen Species/Hypoxia-inducible Factor-1 α /Vascular Endothelial Growth Factor Pathway. *Journal of cardiovascular pharmacology*, 2014; 64(3): 218-227.
86. Liao, P.L., Lin, C.H., Li, C.H., Tsai, C.H., Ho, J.D., Chiou, G.C., Kang, J.J. and Cheng, Y.W., 2017. Anti-inflammatory properties of shikonin contribute to improved early-stage diabetic retinopathy. *Scientific reports*, 2017; 7(1): 44985.
87. Kim J, Kim Cs, Lee Ym, Sohn E, Jo K, Kim J.S. Vaccinium myrtillus extract prevents or delays the onset of diabetes-induced blood-retinal barrier breakdown. *International Journal of Food Sciences & Nutrition*, 2015; 66: 236-242.
88. A. Jariyapongskul, C. Areebambud, S. Suksamrarn, and C. Mekseepralard. Alpha-mangostin attenuation of hyperglycemia-induced ocular hypoperfusion and blood retinal barrier leakage in the early stage of type 2 diabetes rats. *BioMed Research International*, 2015.
89. Yang, L.P., Sun, H.L., Wu, L.M., Guo, X.J., Dou, H.L., Tso, M.O., Zhao, L. and Li, S.M., 2009. Baicalein reduces inflammatory process in a rodent model of diabetic retinopathy. *Investigative*

- ophthalmology & visual science*, 2009; 50(5): 2319-2327.
90. M. E. Mehrabadi, Z. Salemi, S. Babaie, and M. Panahi. Effect of biochanin A on retina levels of vascular endothelial growth factor, tumor necrosis factor-alpha and interleukin-1beta in rats with streptozotocin-induced diabetes. *Canadian Journal of Diabetes*, 2018; 42(6): 639-644.
 91. R. A. Kowluru and M. Kanwar. Effects of curcumin on retinal oxidative stress and inflammation in diabetes. *Nutrition & Metabolism*, 2007; 4(1): 8.
 92. Gupta, S.K., Kumar, B., Nag, T.C., Agrawal, S.S., Agrawal, R., Agrawal, P., Saxena, R. and Srivastava, S. Curcumin prevents experimental diabetic retinopathy in rats through its hypoglycemic, antioxidant, and anti-inflammatory mechanisms. *Journal of Ocular Pharmacology and Therapeutics*, 2011; 27(2): 123-130.
 93. J. Li, P. Wang, J. Ying, Z. Chen, and S. Yu. Curcumin attenuates retinal vascular leakage by inhibiting calcium/calmodulin-dependent protein kinase II activity in streptozotocin-induced diabetes. *Cellular Physiology and Biochemistry*, 2016; 39(3): 196-1208.
 94. D. Pradhan, T. Dasmohapatra, and G. Tripathy. Pharmacognostic evaluation of curcumin on diabetic retinopathy in alloxan-induced diabetes through NF-KB and Brn3a related mechanism. *Pharmacognosy Journal*, 2018; 10(2): 324-332.
 95. Yang, F., Yu, J., Ke, F., Lan, M., Li, D., Tan, K., Ling, J., Wang, Y., Wu, K. and Li, D. Curcumin alleviates diabetic retinopathy in experimental diabetic rats. *Ophthalmic research*, 2018; 60(1): 43-54.
 96. Zuo ZF, Zhang Q, Liu XZ. Protective effects of curcumin on retinal Muller cell in early diabetic rats. *International Journal of Ophthalmology*, 2013; 6(4): 422-424.
 97. Kang, M.K., Lee, E.J., Kim, Y.H., Kim, D.Y., Oh, H., Kim, S.I. and Kang, Y.H. Chrysin ameliorates malfunction of retinoid visual cycle through blocking activation of AGE-RAGE-ER stress in glucose-stimulated retinal pigment epithelial cells and diabetic eyes. *Nutrients*, 2018; 10(8): 1046.
 98. Kang, M.K., Park, S.H., Kim, Y.H., Lee, E.J., Antika, L.D., Kim, D.Y., Choi, Y.J. and Kang, Y.H. Dietary compound chrysin inhibits retinal neovascularization with abnormal capillaries in db/db mice. *Nutrients*, 2016; 8(12): 782.
 99. Zhang L, Zhang ZK, Liang S. Epigallocatechin-3-gallate protects retinal vascular endothelial cells from high glucose stress in vitro via the MAPK/ERK-VEGF pathway. *Genet Mol Res.*, 2016; 15(2): 10-4238.
 100. P. Lv, J. Yu, X. Xu, T. Lu, and F. Xu. Eriodictyol inhibits high glucose-induced oxidative stress and inflammation in retinal ganglial cells. *Journal of Cellular Biochemistry*, 2019; 120(4): 5644-5651.
 101. Wu, J., Ke, X., Ma, N., Wang, W., Fu, W., Zhang, H., Zhao, M., Gao, X., Hao, X. and Zhang, Z. Formononetin, an active compound of *Astragalus membranaceus* (Fisch) Bunge, inhibits hypoxia-induced retinal neovascularization via the HIF-1 α /VEGF signaling pathway. *Drug Design, Development and Therapy*, 2016; 10: 3071-3081.
 102. Dongare S.H., Rajendran S.H., Senthilkumari S., Gupta S.K., Mathur R.A., Saxena, R.O. and Srivastava S. Genistein alleviates high glucose induced toxicity and angiogenesis in cultured human RPE cells. *International Journal of Pharmacy and Pharmaceutical Sciences*, 2015; 7(8).
 103. Xin H, Zhou F, Liu T, Li GY, Liu J, Gao ZZ, Bai GY, Lu H, Xin ZC. Icaritin ameliorates streptozotocin-induced diabetic retinopathy in vitro and in vivo. *International Journal of Molecular Science*, 2012; 13: 866-878.
 104. X. H. Xu, C. Zhao, Q. Peng, P. Xie, and Q. H. Liu. Kaempferol inhibited VEGF and PGF expression and in vitro angiogenesis of HRECs under diabetic-like environment. *Brazilian Journal of Medical and Biological Research*, 2017; 50(3).
 105. Chen Y, Sun XB, Lu HE, Wang F, Fan XH. Effect of luteoin in delaying cataract in STZ-induced diabetic rats. *Arch Pharm Res*, 2017; 40: 88-95.
 106. L. H. Liu, Z. F. Zuo, S. J. Lu, A. H. Liu, and X. Z. Liu. Naringin attenuates diabetic retinopathy by inhibiting inflammation, oxidative stress and NF-kappa B activation in vivo and in vitro. *Iranian Journal of Basic Medical Sciences*, 2017; 20(7): 813-821.
 107. Kim, J., Kim, K.M., Kim, C.S., Sohn, E., Lee, Y.M., Jo, K. and Kim, J.S. Puerarin inhibits the retinal pericyte apoptosis induced by advanced glycation end products in vitro and in vivo by inhibiting NADPH oxidase-related oxidative stress. *Free Radical Biology and Medicine*, 2012; 53(2): 357-365.
 108. Teng Y, Cui H, Yang M, Song H, Zhang Q, Su Y, Zheng J. Protective effect of puerarin on diabetic retinopathy in rats. *Molecular Biology Reports*, 2009; 36: 1129-1133.
 109. X. Wang, H. Li, H. Wang, and J. Shi. Quercetin attenuates high glucose induced injury in human retinal pigment epithelial cell line ARPE-19 by up-regulation of miR-29b. *Journal of Biochemistry*, 2020; 167.
 110. F. Ghadiri Soufi, E. Arbabi-Aval, M. Rezaei Kanavi, and H. Ahmadih. Anti-inflammatory properties of resveratrol in the retinas of type 2 diabetic rats. *Clinical and Experimental Pharmacology and Physiology*, 2015; 42(1): 63-68.
 111. Ola MS, Ahmed MM, Ahmad R, Abuhashish HM, Al-Rejaie SS, Alhomida AS. Neuroprotective effects of rutin in streptozotocin-induced diabetic rat retina. *Journal of Molecular Neuroscience*, 2015; 56: 440-448.

112. Ahmad, S., ElSherbiny, N.M., Jamal, M.S., Alzahrani, F.A., Haque, R., Khan, R., Zaidi, S.K., AlQahtani, M.H., Liou, G.I. and Bhatia, K. Anti-inflammatory role of sesamin in STZ induced mice model of diabetic retinopathy. *Journal of Neuroimmunology*, 2016; 295:47-53.
113. H.T. Zhang, K. Shi, A. Baskota, F.-L. Zhou, Y.-X. Chen, and H.-M. Tian. Silybin reduces obliterated retinal capillaries in experimental diabetic retinopathy in rats. *European Journal of Pharmacology*, 2014; 740: 233–239.
114. Ahiskali, I., Pinar, C.L., Kiki, M., Mammadov, R., Ozbek Bilgin, A., Hacimuftuoglu, A., Cankaya, M., Keskin Cimen, F. and Altuner, D. Effect of taxifolin on development of retinopathy in alloxan-induced diabetic rats. *Cutaneous and Ocular Toxicology*, 2019; 38(3): 227-232.
115. Chung HK, Choi SM, Ahn BO, Kwak HH, Kim JH, Kim WB. Efficacy of troxerutin on streptozotocin-induced rat model in the early stage of diabetic retinopathy. *Arzneimittelforschung*, 2005; 55: 573-580.
116. Z. Yu, B. Lu, Y. Sheng, L. Zhou, L. Ji, and Z. Wang. Andrographolide ameliorates diabetic retinopathy by inhibiting retinal angiogenesis and inflammation. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 2015; 1850(4): 824–831.
117. Dong LY, Jin J, Lu G, Kang XL. Astaxanthin attenuates the apoptosis of retinal ganglion cells in db/db mice by inhibition of oxidative stress. *Mar Drugs*, 2013; 11: 960-974.
118. Dene BA, Maritim AC, Sanders RA, Watkins JB III. Effects of antioxidant treatment on normal and diabetic rat retinal enzyme activities. *Journal of Ocular Pharmacology & Therapeutics*, 2005; 21: 28-35.
119. Cai, Y., Li, W., Tu, H., Chen, N., Zhong, Z., Yan, P. and Dong, J. Curcuminolide reduces diabetic retinal vascular leukostasis and leakage partly via inhibition of the p38MAPK/NF- κ B signaling. *Bioorganic & medicinal chemistry letters*, 2017; 27(8): 1835-1839.
120. Kim SH, Jung SH, Lee YJ, Han JY, Choi YE, Hong HD, Jeon HY, Hwang J, Na S, Kim YM, Ha KS. Dammarenediol-II prevents VEGF-mediated microvascular permeability in diabetic mice. *Phytotherapy Research*, 2015; 29: 1910-1916.
121. Arnal, E., Miranda, M., Johnsen-Soriano, S., Alvarez-Nölting, R., Díaz-Llopis, M., Araiz, J., Cervera, E., Bosch-Morell, F. and Romero, F.J. Beneficial effect of docosahexanoic acid and lutein on retinal structural, metabolic, and functional abnormalities in diabetic rats. *Current eye research*, 2009; 34(11): 928-938.
122. S. Li, H. Yang, and X. Chen. Protective effects of sulforaphane on diabetic retinopathy: activation of the Nrf2 pathway and inhibition of NLRP3 inflammasome formation. *Experimental Animals*, 2019; 68(2): 221-231.
123. T. F. Tzeng, S. S. Liou, Y. C. Tzeng, and I. M. Liu. Zerumbone, a phytochemical of subtropical ginger, protects against hyperglycemia-induced retinal damage in experimental diabetic rats. *Nutrients*, 2016; 88.
124. S. Shanmuganathan and N. Angayarkanni. Chebulagic acid chebulinic acid and gallic acid, the active principles of triphala, inhibit TNF α induced pro-angiogenic and pro-inflammatory activities in retinal capillary endothelial cells by inhibiting p38, ERK and NF κ B phosphorylation. *Vascular Pharmacology*, 2018; 108: 23-35.
125. X. Mei, L. Zhou, T. Zhang, B. Lu, Y. Sheng, and L. Ji. Chlorogenic acid attenuates diabetic retinopathy by reducing VEGF expression and inhibiting VEGF-mediated retinal neoangiogenesis. *Vascular Pharmacology*, 2018; 101: 29-37.
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